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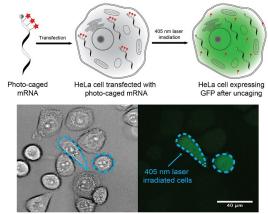
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UC San Diego Chemists Use Light to Pinpoint Gene Expression

Armed with skill, special tools and light, University of California San Diego Associate Professor Neal Devaraj and a group of his chemistry graduate students activated cellular gene expression with unique precision.

By modifying messenger RNA (mRNA)—a group of molecules that carries genetic information from DNA to ribosomes where specifications of gene expression occur—the chemists were able to precisely trigger gene expressions at a specific time and place using laser light. This novel technique will ease future studies of individual protein functions in cells or tissues at different stages of biological development.

The details of this study are published online in a recent issue of <u>Angewandte Chemie</u>.



Spatiotemporal-activation of EGFP expression in HeLa cells

The figure demonstrates the spatiotemporal-activation of cellular gene expression at a single cell level. The cartoon (top) shows the experimental procedure. The cell images show that upon laser irradiation of selected cells, which triggered the cleavage of photo-caging groups, mRNA translation were reactivated within those selected cells. Image courtesy of the Devaraj Research Group.

Scientifically, manipulating gene expression is fundamental to basic research, biotechnology and the development of new therapies. And, according to Devaraj, Department of Chemistry and Biochemistry faculty member and principal investigator at the <u>Devaraj Research Group</u> at UC San Diego, the ability to regulate the translation of mRNA provides a unique way to control the function of novel mRNA-based therapeutics.

"mRNA has recently been widely studied as a drug for therapeutic applications," confirmed Devaraj. "Compared to DNA-based gene therapy, in vivo expression of therapeutic proteins through the direct translation of mRNA offers much quicker response times and minimizes the risk of causing insertional mutagenesis (DNA mutations). Our mRNA translation regulation approach can greatly expand the toolbox of mRNA modification for therapeutic discoveries." Devaraj explained that the study required a dynamic combination of organic chemistry, molecular biology and cell biology. A basic explanation of the process is that scientists produced photo-reactive groups of chemicals using exposure to light and designed RNA sequences by cloning and mutating molecules. To obtain the modified mRNA—of the type that would express mammalian traits—the researchers conducted a variety of chemical reactions using enzymes.

"The obtained mRNA was transfected into cultured HeLa cells, and its translation activity was measured by fluorescence microscopy," described Deveraj, adding that researchers involved in the project were able to learn different synthetic skills as well as molecular biology techniques.

"The most exciting part about chemical biology is that it requires knowledge and application of both chemical and biological techniques. By designing and chemically modifying biomacromolecules, we are able to artificially control different cellular pathways and develop novel techniques which can promote the development of both fundamental types of research and biotechnological applications," said Devaraj.

The research was mainly conducted by UC San Diego graduate students Dongyang Zhang, Cun Yu Zhou and Kayla Busby. Former postdoctoral scholar Seth Alexander also contributed to the study. Funding was provided by the National Institutes of Health (grants R01 GM123285-01 and T32 GM112584-03) and the Army Research Office (grant W911NF-13-1-0383).

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