

SHILLARD

1949

Journal I

Return to

Zoo Drexel

Chicago

5624

Dochester

6 per

Tunwell

~~Dochester~~

Heins

Brain Blood barrier

for idea of cranial nodes properly

See Louis Fleener

Concepte Inst. of Embryology
Baltimore

Julius Hopkins, Medical School Campous

egg

Walter Landauer, Dept of Genetics
Univ. of Conn.
Storrs, Conn.

Leitch, Mrs Claude Tweedle
office Ea 7771 ext 269
Barrack C

egg

Bulletin 262 Apr 1948

The Hatchability of Chicken eggs
as influenced by environment
and Heredity. —

Walter Landauer

Storrs Agricultural Experiment
Station College of Agriculture
Univ of Conn, Storrs, Conn.

Oxygen sol in water at 0°C

~~It~~ $\sim \frac{5}{100}$ of cc per cc

Carbon dioxide $\sim \frac{3}{100} \frac{1}{100}$ cc/cc in
equilibrium with atmospheric air
Algae growing once in 20 hours
compared with chemostat at $\tau = 2$ hours
would be 20 times more of at equal
water (variable amount) but may be 10 times
better at equal ~~water~~ dry substance
per cc ~~but~~ 4 times better of because of
 $\frac{1}{4}$ efficiency of Cali. — ~~5 times more of~~
since oxygen in air is ~~that rate~~ algae

or 40 ~~times~~ $\frac{40}{4} = 10$ times dry substance $\frac{40}{4} = 10$

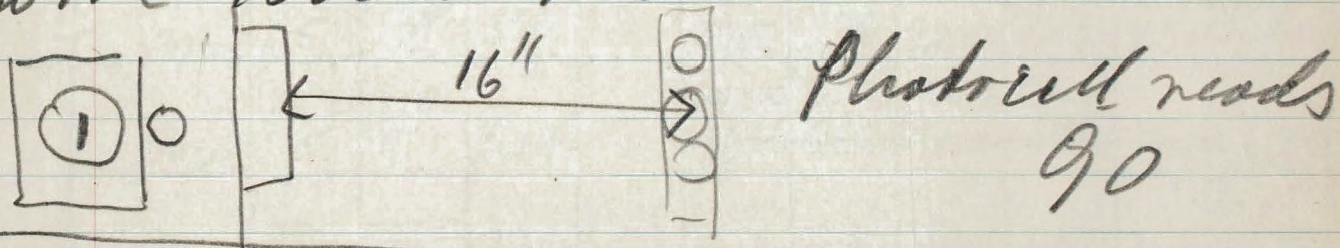
Exp on B/r UV irradiation
- light reaction -

In these experiments B/r was
grown in F ^{to ~ 10⁸} ~~at 10⁸~~ transferred
to saline and left in saline
overnight or longer; alternatively
it was irradiated in F while
growing (water around 510⁸). -

UV lamp from bulb to shaker
plate 50 cm, (Photocell reads
about 50)

UV irradiation in petri dish,
6 inches, and 20cc is put in

Spot light standard procedure
with 1000 Watt bulb



Graph No 1 shows inactivation
curves for B/r
in saline and F

Effect of UV and Light on mutants

First experiment on B/r phage resistant mutants: See Graph 2, Exp of Apr 2nd 1949

2.5 min UV on B/r resting in saline regrown and plated

for multiplication of:
500 in dark we have $24000/10^8$ (T₁)
400 in light $5000/10^8$ (T₁)

* corrected for parent mutants
T₇/T₁

| |
|--------|
| 5* |
| 5 |
| 4.5 |
| 4* |
| 4 |
| 6 |
| approx |
| 2 |
| 2.5 |

Double mutants with UV

B/sr
4/22/49 UV 0, 30, 50, 80 sec UV
Dark and light
Mutants after regrow
1:20 dilution before regrow

Mutants Dark
Mutants Light

B/r Graph 3

| | | | |
|----|----------------|----------------|----------------|
| | T ₁ | T ₆ | T ₇ |
| 30 | 3.0 | 5.1 | 4.4 |
| 50 | (5.6) 2.8 | (4.8) 2.4 | (7.0) 3.5 |
| 80 | 9.8 | 9.4 | 8 |

| | UV | $\times 10^7$ Essay | Regrow Assay | Multipl Gen | T ₁ | T ₁ /10 ⁸ | T ₆ | T ₆ /10 ⁸ | T ₇ | T ₇ /10 ⁸ |
|---|--------|---------------------|--------------|-------------|----------------|---------------------------------|----------------------|---------------------------------|----------------|---------------------------------|
| | 0 | 10 | 310 | 600 | 2000 | 64.5 | 1100 | 35.5 | 5700 | 184 |
| D | 30 sec | 6 | 120 | 400 | 10,000 | 835 | 10,000 | 835 | 45,000 | 3750 |
| D | 50 sec | 4.5 | 130 130 | 560 | 28,000 | 2150 | 20,000 | 1540 | 135,000 | 10,400 |
| D | 80 sec | 2.2 | 100 100 | 880 | 66,000 | 6600 | 63,000 | 6300 | 320,000 | 32,000 |
| | 0 | 8 | 110 130 | 225 | 800 | 88 | — | — | — | — |
| L | 30 sec | 8.5 | 135 135 | 320 | 3700 | 274 | 2200 | 163 | 11,500 | 850 |
| L | 50 sec | 7.5 | 110 110 | 294 | (4200) 8600 | (382) 780 | (3500) 7000 | (320) 640 | 33,000 | (1500) 3000 |
| L | 80 sec | 6.5 | 75 | 230 | 5000 | 670 | 5000 | 670 | 30,000 | 4000 |
| | approx | 0 | 11 | — | — | — | — | — | — | — |
| D | 80 | 2.3 | 70 | 600 | 56,000 | 8000 | 2600 | 10600 | 106000 | 15000 |
| L | 80 | (5)? | 46 | (180)? | 4760 | 1000 | 2.5% 2.5% 2.5% | 11700 | 2500 | 2500 |

| 4-26-49 | Assay before UV | Assay after UV | Regraw Assay | Multiplication | T1 | T1/10 ⁸ | T7 | T7/10 ⁸ | (T7/T1) |
|------------------------|----------------------|---------------------|--|----------------|------------------|--------------------|--------------------|--------------------|---------|
| 40 F | 1.15x10 ⁸ | 3x10 ⁷ | 1.6x10 ⁹ | 500 | 50,000 | 3100 | 150,000 | 9400 | 3 |
| 40 S | 6.3x10 ⁷ | 4.4x10 ⁷ | 9.8x10 ⁹ | 2300 | 92,000 | 920 | 274,000 | 2800 | 3 |
| ± 80 F | 1.15x10 ⁸ | 2x10 ⁶ | 1.5x10 ⁸ 2x10 ⁶ | 750 1000 | 16,300 18,200 | 11,000 9100 | 34,200 34,400 | 22,800 17,200 | 2 |
| 80 S | 6.3x10 ⁷ | 2.2x10 ⁷ | 7x10 ⁹ | 3200 | 372,000 | 5330 | 800,000 | 11400 | 2 |
| 80 S (B ₂) | 6.3x10 ⁷ | 2.2x10 ⁷ | 1.8x10 ⁹ | 820 | 97,000 | 5450 | 500,000 | 27800 | 5L !! |

Vary I: for fixed UV

5/2r graph 4 strains for 1 hrs H
light in 37° paper incubator
by light after 3 min incubation
(water drop from 10⁹ cc to 650 pcc)
for different light intensities. —

~~Saturday Apr 30th~~

M

Experiment with back-mutation
of threonineless (W) strain
on Apr 25th/49, since UV dropped when
from 1.25×10^7 to 1.5×10^6 giving by
removes plating with 0.003 mg/ml threonine
on plate 3560 mutants/c [same with ten times
more threonine]

This would give $\frac{3560 \times 100}{1.5} / 10^8 = 240,000 / 10^8$

with no U.V on 0.01 mg/ml plate 2083 colonies
[at least]

B/r UV series, reprints for pore² law

| UV | Assay | Regrow dist | Regrow dist | Regrow dist | Regrow assay | Multipl | T _{1/10} | Con T _{1/10} | T _{6/10} | Con T _{6/10} | Multipl | Multipl | Con T _{4/10} | |
|--------|-------------------------------|-------------|------------------------|------------------------|--------------------------|---------|-------------------|-----------------------|-------------------|-----------------------|--------------------|---------|-----------------------|--------|
| 0 | 1.3 x 10 ⁸ | 5 100 | 4 | 3 20 | 2.9 x 10 ⁸ | 135 | 70 | 70 | 30 | 30 | | 155 | 370 | |
| 15 | 1.2 x 10 ⁸ | 1 20 | 1 | 4 20 | 3.4 x 10 ⁸ | 265 | 164 | 94 | 162 | | | 265 | 1280 | |
| 30 | 1.1 x 10 ⁸ | 1 20 | 1 | 12 20 | 4.6 x 10 ⁸ | 270 | 610 | 540 | 542 | | | 290 | 3790 | |
| 80 | 4.6 x 10 ⁷ | 1/2 20 | 1 | 1 20 | 3.2 x 10 ⁸ | 270 | 6000 | 6000 | 6000 | | | 270 | 55,000 | |
| 160 | 6.9 x 10 ⁴ | 10 100 | 1 | 1 | | | | | | | | 850 | 175,000 | |
| Repeat | | | | | | | | | | | | | | |
| 0 | | | | | | | | | | | | | | |
| 80 | 3.14 x 10 ⁷ WWD | | | | 1.2 x 10 ⁹ | 1500 | 7200 | | 58000 | | | 1000 | 1500 | 25000 |
| 160 | 5.57 x 10 ⁴ WWD | | | | 4.4 x 10 ⁶ | 800 | 32500 | | 28000 | | | 165 | 800 | 126000 |

B/r
Graph 5

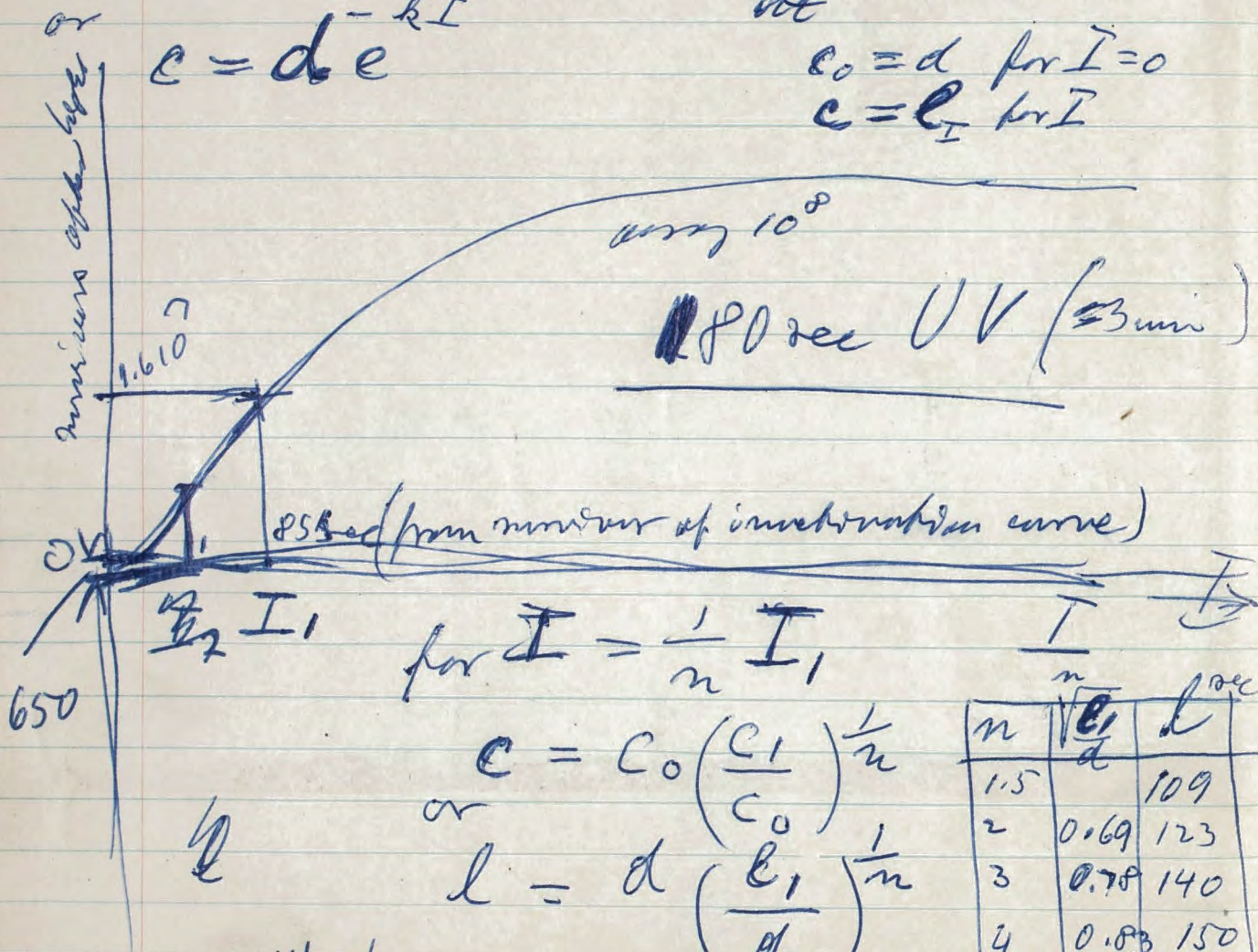
A certain amount of chemical d is produced by the UV irradiation. When we shine light of intensity I for 1 hour we have (writing $c_0 = d$)

$$-\frac{dc}{dt} = kIc$$

$$c = d e^{-kI}$$

$$c_0 = d \text{ for } I=0$$

$$c = c_I \text{ for } I$$



$$\text{for } I = \frac{1}{n} I_1$$

$$c = c_0 \left(\frac{c_1}{c_0} \right)^{\frac{1}{n}}$$

$$\text{or } d = d \left(\frac{d_1}{d} \right)^{\frac{1}{n}}$$

| n | $\frac{c_1}{c_0}$ | d |
|---------------|-------------------|-----|
| 1.5 | | 109 |
| 2 | 0.69 | 123 |
| 3 | 0.78 | 140 |
| 4 | 0.83 | 150 |
| 5 | 0.86 | 155 |
| 6 | 0.885 | 159 |
| 8 | 0.915 | 164 |
| $\frac{1}{2}$ | | 40 |
| 1 | | 85 |

$$I k = \ln \frac{d}{c}$$

$$k = \frac{1}{I} \ln \frac{d}{c}$$

$$\text{for } \begin{cases} n = \infty \\ c = d \end{cases}$$

| UV | Assay | after light | Rayon Blanch | Rayon Assay | Multiple | $T_1/10^8$ | $T_6/10^8$ | $T_4/10^8$ | 2 | $T_1/10^8$ | $T_6/10^8$ | $T_4/10^8$ |
|---------------|-------------------|-------------------|-----------------|--------------------|----------|------------|------------|------------|-------|------------|------------|------------|
| 0 | 1.3×10^8 | | | | | | | | | | | |
| 220 | | | | | | | | | | | | |
| 270 | — | 1.3×10^6 | 1:15 | 5.36×10^7 | 600 | 15,400 | 10,500 | 68,500 | 12620 | 16,000 | 16,000 | 110,000 |
| | | | | | | Exp | | | | Theory | | |

This experiment to show that rolling after light reactivation also determines mutants correctly. Mutants interpolated or extrapolated with "square of dose" law to get theoretical values. —
 For interpolation see graph (B/r) 6

4.) Use a fixed UV say 3 min and 1 1/2 min
and give for 1 hr light at varied
intensities

measure: mutants

measure: mutants

a) see if mutants correspond to method
those 1 (for 3 min UV)

b) see if both mutants and mutants
vary with same factor \downarrow of light
intensity for the two chosen UV doses
above (3 min 1 1/2 min); of this the
mutants will be a more sensitive
test for ~~strong~~ strong lights and
the mutants will be a more sensitive
test for small recombination, particularly
for small recombination in 3 min
U.V. case

5.) Control death by light alone (full strength)

6.) How does ^(full strength) reactivation depend on duration
of light say for 270 sec UV
^{upto 3 hrs} (best for mutants)

7.) Take 30 sec UV reprogram, use as
starting material: give it 160 sec UV
reprogram best for mutants

Theoretical rewrite, —

4

0.) UV series, No light, measure % survivors, go to ~~last~~ ~~dist~~ ~~3/4~~ ~~series~~ ^{500 sec}
as a function of dose d (sec)
Use points 30 sec, 80 sec, 120 sec, 160 sec

1) UV, (no light), replot, measure % mutants [to establish square of dose law] ²⁰⁰
go to 160 sec UV, any further? ²²⁰

2) UV + 1 hr light, ~~measure~~ measure % survivors, to determine whether ~~the~~ ^{mutant} dose law ~~is~~ defined by survivors from curve established under 0.) is equal to Q (about 2.5) $\times d$ (sec)
go out to $3\frac{1}{2} \times 2.5$ min
 $230 \text{ sec} \times 2.5 = 570 \text{ sec}$

3.) UV + 1 hr light replot measure % mutants, — go out to 270 sec UV and beyond?
~~400 sec (T1) to dose 10 min/sec~~
~~450 sec (T4) " " " "~~
From 1.) and 3) get ~~ratio~~ Q def " back "
This should be done now for 30, 80, 160 sec UV. " light "
From 2 and 3 one would see whether mutants correspond to dose d calculated from survivors

P. 1) Delayed appearance of new bands
in serum

9. 1) Much lower ratio

Theory of light reabsorption at
 rooted light intensity: (Durostom)
 What about experiments of this type:
 a UV dose is given and non recoverable
 death is measured as a function of
 time between UV dose and light
 application a) how does this depend on
 the U.V. dose? b.) if some light is given
 immediately after UV and then we
 incubate for varying periods of time
 before we plate.)

Theory: assume there is a deplorable
 which is some function of the
 concentration (c) of the poison c
Question: what does it mean? If light leaves
 $\frac{1}{4}$ of mutants, or $\frac{1}{10}$ of mutants (at higher UV
 dose)
 it means it leaves $\frac{1}{2}$ of or $\frac{1}{3.3}$ of poison
 Why should it remove a larger fraction
 of the poison at higher U.V. doses??
 Perhaps because the latter one is a
deeper lay!

Effect of filler 73P on reactor output

160 sec UV irradiated from $[1.34 \times 10^8 \text{ P}]$
down to $[22,400]$ or drop from 10^8
to $54,000$.

1 hr light Magpul 1000 watt at 8 inches
with cell holder (see before) reactivated
without filler 5.94×10^7
with filler [molded] 5.61×10^7
or 5% less within exp error.

see back III Mag 10/49

Holding by light

H

B/r in saline for $2\frac{1}{2}$ days
with Magul 1000 Watt lamp at
8 inches [with $\frac{5}{100}$ normal cullr in
one dish 100 pyrex flasks]

ones in holding in 37° room
with fans on [temp rise estimated
at 2°C]

see Book III May 10 149

Light calibrations

~~with Hg Lamp~~

Points from 1000 Watt lamp
(120 Volt Magul) to photo cell line
pane with hole I in Murchell No 2
with filter 9883 59.5 down stairs
" " B580 50 - down stairs

Hg lamp goes up (close by) to 200
down stairs with 9883
and with 9883 + 738 it goes to 70. -
738 cuts to $\frac{1}{3}$ rd

To get ~~light~~ dark to light
 ratio at ~~180 sec~~ 76 second
 extrapolate back from light
 from light 180 second with square law

| | | | |
|--------------------|----------------|----------------|-----------------|
| D 76 sec dark | T1/108 6700 | T6/108 6000 | T7/108 25000 |
| L 76 sec extrapol. | 1600 | 1140 | 5000 |
| | 4.2 | 5.25 | 5 |
| | 2.1 | 2.3 | 2.24 |

These figures have to be compared
 to 2.51

If we extrapolate ~~to~~ the T6 case from
 180 sec 7100 with square law } $\frac{175 \text{ sec}}{76 \text{ sec}} = 2.3 = q$
 to 175 sec ~~6000~~

similarly:

| | |
|------------------------|---|
| for T1 case $q = 2.05$ | } This is excellent agreement with above values (2.1, 2.3, 2.24) which uses square law very essentially |
| for T6 case $q = 2.3$ | |
| for T7 case $q = 2.24$ | |

See graph 12

UV assay Regions Assay after Regions Multi Corn T₁/10 Corn T₆/10 Corn T₄/8 SIZE SAMPLE T₁ T₄/T₁ T₄/T₆

low right

| | | | | | | | | | | |
|-----|----------------------|-----------|---------------------|------|--------|--------|--------|-----|------|-----|
| 0 | 8.1x10 ⁷ | 100 | 9.7x10 ⁸ | 1200 | 53 | 29 | 210 | | 4 | 7.2 |
| 30 | 5.4x10 ⁷ | 80 | 10 ⁹ | 1480 | 766 | 506 | 2550 | 400 | 3.3 | 5.0 |
| 76 | 2.1x10 ⁷ | 20 | 1.4x10 ⁹ | 1290 | 6700 | 6000 | 25000 | 670 | 3.7 | 4.2 |
| 76 | " | 200 | — | | | | | | | |
| 114 | 3.06x10 ⁶ | 20 | 2.1x10 ⁸ | 1370 | 11,900 | 14,600 | 66000 | 400 | 5.5 | 4.5 |
| 152 | 5.72x10 ⁴ | 10 | 5.2x10 ⁶ | 920 | 35000 | 16000 | 110000 | 200 | 3.2 | 6.9 |
| 170 | 6.34x10 ³ | No REGROW | | | | | | | | |
| 190 | 2.35x10 ⁷ | 40 | 6.4x10 ⁸ | 900 | 9700 | 7100 | 31000 | 990 | 3.1 | 4.4 |
| 285 | 3.2x10 ⁶ | 20 | 2.7x10 ⁸ | 1400 | 11,700 | 10,700 | 44,300 | 300 | 3.8 | 4.1 |
| 380 | 8.5x10 ⁴ | 10 | 9.3x10 ⁶ | 1000 | 14,900 | 5500 | 41000 | 120 | 2.75 | 7.4 |
| 425 | 8.26x10 ⁵ | 10 | 8.3x10 ⁵ | 1000 | 23,500 | 18,000 | 97,500 | 20 | 4.2 | 5.4 |
| 475 | 613 | No REGROW | | | | | | | | |
| 551 | 20 | " " | | | | | | | | |

must be direct because it falls off curve

low left

FIRST Measurement

| | | | | | |
|-----|----------------------|------|-------|-------|-------|
| 170 | 5.06x10 ⁸ | 860 | 10800 | 8500 | 36800 |
| 175 | 2.45x10 ⁸ | 1520 | 9600 | 7000 | 40700 |
| 380 | 7.15x10 ⁸ | 835 | 12700 | 7700 | 47000 |
| 425 | 5.2x10 ⁵ | 600 | | 28000 | |

B for March May 13th/49, May 11th/49
for square law see graph No 7

This is in graph No 10
No 11

gives $\alpha = 2.51$

$$\frac{L_n}{D} = \frac{1}{q} + \left(1 - \frac{1}{q}\right) \left(\frac{L_0 - \frac{1}{q}}{1 - \frac{1}{q}}\right)^n$$

q can be determined for measuring
for a given D the value of L_0 and
another value L_n —

Theory of inductively coupled effect

For infinite inductivity and any 2 turns
we find dose L (after by hit) = $\frac{D}{Q}$

we may then write

$$L = \frac{D}{Q} + \left(1 - \frac{1}{Q}\right) D e^{-\alpha I t}$$

For instance if we give a dose of 180 sec
and of $Q = 2.5$

$$L = \frac{180}{2.5} + \left(1 - \frac{1}{2.5}\right) 180 e^{-\alpha I t}$$

we now determine ~~the value of~~ L for
various values of I . - If we find

$L = L_0$ for $I = I_0$

$$L_0 = \frac{180}{2.5} + \left(1 - \frac{1}{2.5}\right) 180 e^{-\alpha I_0 t}$$

$$\frac{L_0 - \frac{180}{2.5}}{180 \left(1 - \frac{1}{2.5}\right)} = e^{-\alpha I_0 t} = \frac{L_0 - \frac{D}{Q}}{D \left(1 - \frac{1}{Q}\right)} = e^{-\alpha I_0 t}$$

For example for $I_0 = 22.5$ $L_0 = 120$ sec
 $e^{-\alpha I_0 t} = \text{almost } 0.445$

for $I = 2I_0$ $L = 72 + 108 (0.445)^2 = 93.4$
= 45

for $I = 4I_0$ $L = 72 + 108 (0.445)^4 = 72 + 108 \times 0.04$
= 90

$$L = \frac{D}{Q} + \left(1 - \frac{1}{Q}\right) D e^{-\alpha I t}$$

for $I = n I_0$

newbie

~~Q~~ relates to no delay

q_1 relates to one hour delay

q_t relates to t hour delay

~~q_n~~ q_n meant q_t for $t = n$ hours
 $n = 3$

$$\frac{q_{n-1}}{q_n} \cdot \frac{Q-1}{Q} = \left(\frac{q_1-1}{q_1} \cdot \frac{Q-1}{Q} \right)^n$$

$$\frac{Q-1}{Q^{n+1}} \left(\frac{q_{n-1}}{q_n} \cdot \frac{Q-1}{Q} \right)^{\frac{1}{n}} = \frac{q_{n-1}-1}{q_{n-1}} = \frac{0.9}{1.9} = 0.475$$

$$0.412 \cdot \frac{q_{n-1}-1}{q_{n-1}} \cdot \frac{Q-1}{Q} = \left(\frac{q_{n-1}-1}{q_n} \cdot \frac{Q-1}{Q} \right)^{\frac{n-1}{n}} \cdot \frac{Q-1}{Q}$$

Exp at May 18 $11/13$

- $q_3 = 1.53$
- $q_2 = 1.90$
- $q_1 = 1.90$
- $Q = 2.14$

Notch equation: $0.396 m$

$$\frac{q_{m-1}}{q_m} = \left(\frac{q_3-1}{q_3} \cdot \frac{Q-1}{Q} \right)^{\frac{3}{m}} \cdot \frac{Q-1}{Q}$$

for instance $m=6$

$$\frac{q_{6-1}}{q_6} = \left(\frac{0.53+2.14}{1.53 \cdot 2.14} \right)^{\frac{2}{1.14}} \cdot 2.14$$

Theory of Time Delay effect:

H

We assume that restorable portion $D(1 - \frac{1}{q})$ decays exponentially with the same λ .
 Then after delay of t hours we have a dose L

$$L = \frac{D}{q} + \left\{ (1 - \frac{1}{q})D - (1 - \frac{1}{q})D e^{-\lambda t \text{ delay}} \right\}$$

$$L_t = D - (1 - \frac{1}{q})D e^{-\lambda t \text{ delay}}$$

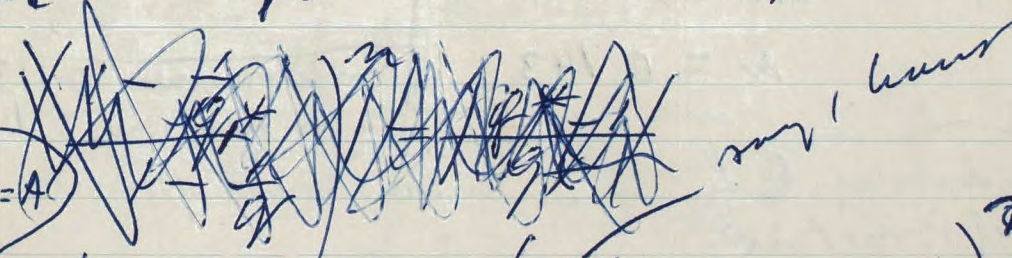
$$\frac{L_t}{D} = 1 - (1 - \frac{1}{q}) e^{-\lambda t \text{ delay}}$$

$$\frac{L_t^*}{D} = \frac{1}{q^*}$$

$$\frac{1}{q^*} = 1 - (1 - \frac{1}{q}) e^{-\lambda t^*}$$

$$\frac{1 - \frac{1}{q^*}}{1 - \frac{1}{q}} = e^{-\lambda t} = \frac{q^* - 1}{q^*} / \frac{q - 1}{q}$$

If number of hours is n to apply and q_0^* is value of q^* for t_0 hour



$$\frac{q_0^* - 1}{q_0^*} / \frac{q - 1}{q} = \left(\frac{q_0^* - 1}{q_0^*} / \frac{q - 1}{q} \right)^n$$

2.26
 7.74
 1.29 1.14 = A
 3.04 2.14
 1.135
 1.745
 A = 0.0226
 q_6 A = q_6 - 1 or q_6 / (A - 1) = -1 4.17 - 1.135 = 1.029

Because we assume

$$x_0 = a \text{ Dose}$$

$$y_0 = b \text{ dose}$$

$$\text{where } a + b = 1$$

$$\frac{L}{\text{Dose}} = a + b \left\{ \frac{k}{\alpha I + \lambda} + \frac{\alpha I}{\alpha I + \lambda} e^{-(\alpha I + \lambda)t} \right\}$$

This is ~~not~~ obviously independent of dose -

$$a = \frac{1}{Q} \quad b = \left(1 - \frac{1}{Q}\right)$$

Value of k could be taken from

3 hour delay experiment

$$Q = 2.14$$

$$\left(\frac{L}{D}\right)_{3 \text{ hours}} = \frac{1}{1.5} = 0.654$$

$$e^{-k \cdot 3} = \frac{0.356}{0.533} = 0.650$$

$$\frac{0.467}{0.533}$$

$$3k = 0.430$$

$$k = 0.143$$

$$L = D - \left(1 - \frac{1}{Q}\right) D e^{-kt}$$

$$\left(\frac{L}{D}\right)_{\text{for 3 hours exp}} = 1 - \left(1 - \frac{1}{Q}\right) e^{-k \cdot 3}$$

$$e^{-k \cdot 3} = \frac{1 - \left(\frac{L}{D}\right)_{3 \text{ hours}}}{1 - \frac{1}{Q}}$$

1 hr delay gives 0.113

2 hr delay gives 0.146

For calculating $\frac{1}{Q}$ from measured point of value of Bracket

$$\frac{\frac{L}{D} - \text{Bracket}}{1 - \text{Bracket}} = \frac{1}{Q}$$

Combined theory of insensitivity
to time delay effects.

Let y be the value of y at time $t=0$
when we start light irradiation
and let us assume that reactance y
~~the~~ amount of prison y changes

$$\frac{dy}{dt} = -y(\alpha I + \nu)$$

$$\text{or } y = y_0 e^{-(\alpha I + \nu)t}$$

When reactance y is present remaining
prison

$$\frac{dx}{dt} = \nu y$$

$$L = x_0 + \int_0^t \nu y dt + y_0 e^{-(\alpha I + \nu)t}$$

$$y_0 \frac{\nu}{\alpha I + \nu} (1 - e^{-(\alpha I + \nu)t})$$

$$L = x_0 + y_0 \frac{\nu}{\alpha I + \nu} (1 - e^{-(\alpha I + \nu)t})$$

~~$$L = x_0 + y_0 \left(\frac{\nu}{\alpha I + \nu} + \frac{\alpha I}{\alpha I + \nu} e^{-(\alpha I + \nu)t} \right)$$~~

$$L = x_0 + y_0 \left\{ \frac{\nu}{\alpha I + \nu} + \frac{\alpha I}{\alpha I + \nu} e^{-(\alpha I + \nu)t} \right\}$$

To calculate λ from a 3 hr delay
(with high intensity) $I = 2I_0$

$$L_{\text{no delay}} = X_0 + y_0 \frac{\lambda}{dI + \lambda}$$

$$L_{\text{3 hours delay}} = X_0 + y_0 + y_0 e^{-\lambda 3} \left(\frac{\lambda}{dI + \lambda} - 1 \right)$$

$$= X_0 + y_0 - y_0 \frac{dI}{dI + \lambda} e^{-\lambda 3}$$

$$= X_0 + y_0 \left(1 - \frac{dI}{dI + \lambda} e^{-\lambda 3} \right)$$

$$= X_0 + y_0 \left(\frac{\lambda + dI(1 - e^{-\lambda 3})}{dI + \lambda} \right)$$

$$= X_0 + y_0 \frac{dI}{dI + \lambda} + y_0 \frac{dI}{dI + \lambda} \left[\lambda 3 - \frac{(\lambda 3)^2}{2} \right]$$

$$L_{\text{3 hours delay}} - L_{\text{no delay}} =$$

$$= y_0 \frac{dI}{dI + \lambda} \left[\lambda 3 - \frac{(\lambda 3)^2}{2} \right]$$

and

$$y_0 = (L_{\text{no delay}})$$

$$L_{\text{no del.}} = X_0 + (1 - X_0) \frac{\lambda}{dI + \lambda}$$

$$L_{\text{no del}} \frac{\lambda}{dI + \lambda} = X_0 \left(\frac{dI}{dI + \lambda} \right)$$

$$X_0 = \frac{L_{\text{no del}} \frac{\lambda}{dI + \lambda}}{\frac{\lambda}{dI + \lambda}} \quad \Bigg| \quad 1 - X_0 = 1 - \frac{L_{\text{no del}} \frac{\lambda}{dI + \lambda}}{\frac{\lambda}{dI + \lambda}}$$

Pre-processor experiment:

M.

~~covering two days. Bugs 13/10 taken from saline suspension on first day bugs were in saline at 3.30 overnight.~~

~~Exp. start on May 10 (Photos read May 11)
Bugs in saline since evening of May 6th.
in ~~the~~ see box, -~~

~~UV inactivation curve
and reconstruction by beam light
at pH given in Graph 13
this shows L/D straight line slope 2.51~~

$$L = x_0 + \int_0^t y dt + y_0 e^{-(dI+k)t}$$

$$\frac{dy}{dt} = -ky$$

back for 3 hours ~~at~~
 $t=0$ to $t=3$ $y = y_0 e^{-kt}$

$$L = x_0 + \frac{y_0}{k} [1 - e^{-k \cdot 3}] + y_0 e^{-k \cdot 3}$$

$$\frac{k}{dI+k} + \frac{dI}{LdI+k} e^{-(dI+k)(t-3)}$$

$$I_{\text{no delay}} = X_0 + y_0 - y_0 \frac{L I}{L I + L}$$

from this

$$e^{-L I} = \frac{1 - \frac{L I_{\text{3h delay}}}{D}}{1 - \frac{L I_{\text{no delay}}}{D}}$$

Assuming that $e^{-L I}$ can be neglected which is true for $I = 2 T_0 (\rho^q)$

M

~~$y_0 = 1 - x_0 =$~~

~~$$\frac{kI}{\alpha I + k} - L + \frac{k}{\alpha I + k} = (\alpha I + k)$$~~

~~$$\frac{\alpha I + k}{\alpha I} = \frac{L_{no\ delay}}{\alpha I} = y_0$$~~

~~$L_{3\ lines} - L_{no\ delay} =$~~

~~$$= \left[1 - \frac{L_{no\ delay}}{\alpha I + k} \right] \left[k^3 - \frac{(k^3)^2}{2!} + \dots \right]$$~~

Repeat

~~$L_{3\ lines\ delay} = \int_0^{\infty} y_0 e^{-kz} \left(\frac{k}{\alpha I + k} \right)^3 dz$~~

~~$$= x_0 + y_0 - y_0 \frac{kI}{\alpha I + k} e^{-kz}$$~~

~~$$= D - y_0 \frac{\alpha I}{\alpha I + k} e^{-kz}$$~~

~~$L_{no\ delay} = x_0 + y_0 \frac{k}{\alpha I + k}$~~

~~$$e^{-kz} = \frac{1 - \frac{L_{delay}}{D}}{1 - \frac{L_{no\ delay}}{D}}$$~~

~~$$1 - \left[1 - \frac{y_0 \frac{\alpha I}{\alpha I + k} e^{-kz}}{D} \right]$$

$$1 = \left[\frac{x_0}{D} + \frac{y_0 k}{D (\alpha I + k)} \right]$$~~

$h(L - L_p)$ for $I=1$ and $I=2$
 ~~$I=2$~~

~~Types 23 for $I=1$ UV 150 and 250~~

June 13 $I=1$ and $I=2$ ~~UV 100~~ $\frac{100}{42} = 2.4$

temp 40°C in $I=2.4$ pos.

half lobes 9 min and 15.8 min
looks like some sub-subsonic

UV = 175

June 9

at $I=2.4$

gives 8.5 min half lobe

June 23 $I=1$ and $I=2$ (Alpha)

$I=1$ 15.8 min (half lobe) $T = 3$ min

$I=2$ 9.5 min $T = 1$ min

at UV = 250

and

$I=1$ for UV = 150

gives half lobe 15.8 min

Room 35°

For write up

(M)

$$10) \int_2(D) = \int_1\left(\frac{D}{Q}\right)$$

May 11

June 19, 21, 23, 25 (Log 20 to 100)

misses in June 16 and 18 test tubes
but

half - life) for $I=1$ and $I=2$

June 23 | June 13 half life 9 min | more good
~~250, 150~~ $I=1$ | $I=1$ and 2:0 | for $I=2$ 2:0
and 16 min. ~~at~~ June 17

indep of d from UV dose

better expressed as Dose rate due
to using D^* for short exp 25 min
reproduction

June 23/49

different sensitivity: say N_a genes
 have r_a form part of the chromosome
 same which has r_a loci
 N_b genes from rest of chromosome
 which has r_b loci then

$$\frac{B}{B_0} = \left[1 - \left(1 - e^{-\frac{r_a}{N_a}} \right)^{N_a} \right] \left[1 - \left(1 - e^{-\frac{r_b}{N_b}} \right)^{N_b} \right]$$

for large r_a and r_b

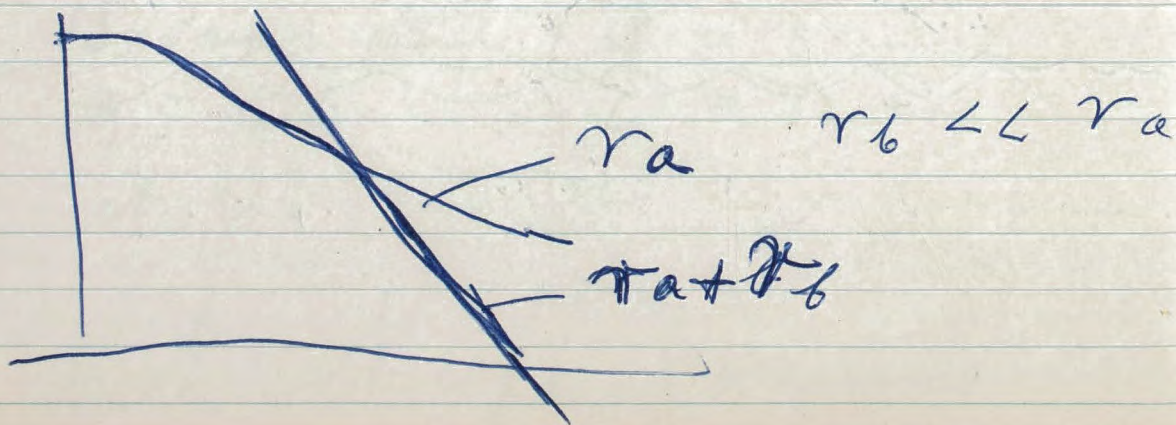
$$\frac{B}{B_0} = \left[2e^{-\frac{r_a}{N_a}} \right]^{N_a} \left[2e^{-\frac{r_b}{N_b}} \right]^{N_b}$$

and

$$\begin{aligned} \ln \frac{B}{B_0} &= N_a \ln 2 - r_a + N_b \ln 2 - r_b \\ &= \underbrace{(N_a + N_b)}_N \ln 2 - \underbrace{r_a + r_b}_r \end{aligned}$$

where r is number of loci on
 chromosome

N total number of genes



Kellum - Brewer

11

Shape of survivor curves

6. Exponential case (equal survivorship of all genes, ~~with~~ and both chromosomes)

Probability of loss of
Number of bits per chromosome r
Number of genes N

probability of survival of one gene $e^{-\frac{r}{n}}$
" " within one gene $1 - e^{-\frac{r}{n}}$
" " " both genes $(1 - e^{-\frac{r}{n}})^2$
" " (escape ~~is~~ ^{variable}) $1 - (1 - e^{-\frac{r}{n}})^2$

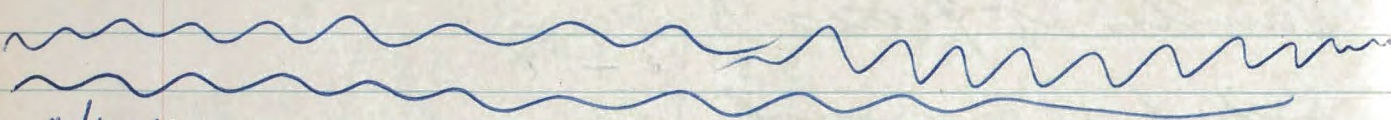
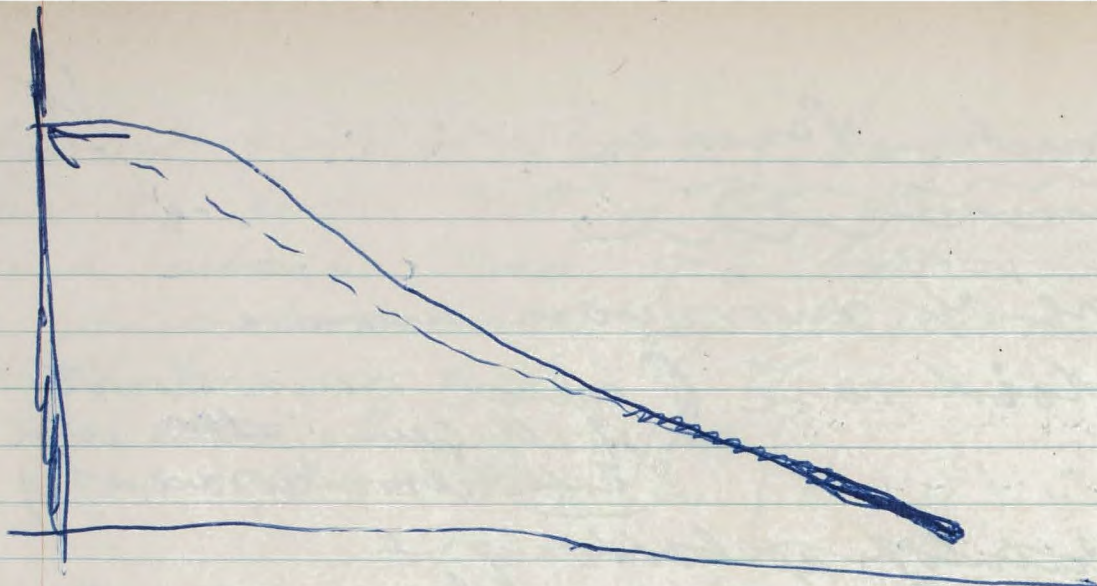
for N genes probability of variable escape

$$\frac{B}{B_0} = \left[1 - (1 - e^{-\frac{r}{n}})^2 \right]^n$$

for large r and $\frac{B}{B_0} \sim \left[2e^{-\frac{r}{n}} \right]^n = 2^n e^{-r}$

$$\ln \frac{B}{B_0} = n \ln 2 - r$$

for $r=0$ extrapolates to $n \ln 2$



~~of~~ N_2 genes require two chromosomes
 and N_3 genes require three
 chromosomes. r_2 looks on that
 part of chromosome which contains
 the N_2 genes; r_3 looks on that part
 of the chromosome that ~~require~~ contain the

N_3 genes;

$$\begin{aligned}
 \frac{B}{B_0} &= \left[1 - \left(1 - e^{-r_1/N_1} \right)^2 \right]^{N_1} \left[1 - \left(1 - e^{-r_2/N_2} \right)^2 \right]^{N_2} \\
 &= \left[1 - \left(1 - e^{-r_1/N_1} \right)^2 \right]^{N_1} \left[1 - \left(1 - e^{-r_2/N_2} \right)^2 \right]^{N_2}
 \end{aligned}$$

for large r_1/N_1 ; r_2/N_2
 $B/B_0 \approx 2^{N_1} e^{-r_1} \times 3^{N_2 - r_2} e^{-r_2}$

equal sensitivity of all genes, but different sensitivity of the two chromosome sources. — I and II

Survival of a gene **I** if chromosome has r hits

probab of ~~survival~~ survival of **I** $e^{-\frac{r}{N}}$

probability that **I** is hit $1 - e^{-\frac{r}{N}}$

probability that

two sources of one is hit $(1 - e^{-\frac{r}{N}}) e^{-\beta \frac{r}{N}}$
 { where βr are hits on }
 chromosome **II**

stable of either ~~one~~ **I** sources

or if ~~one~~ is hit but **II** survives

$$\begin{aligned} \frac{B}{B_0} &= \left[e^{-\frac{r}{N}} + (1 - e^{-\frac{r}{N}}) e^{-\beta \frac{r}{N}} \right]^N \\ &= \left[e^{-\frac{r}{N}} + e^{-\beta \frac{r}{N}} - e^{-(1+\beta)\frac{r}{N}} \right]^N \end{aligned}$$

for large r and $\beta \ll 1$

$$\begin{aligned} \frac{B}{B_0} &\approx \left[e^{-\beta \frac{r}{N}} \right]^N \approx e^{-\beta r} \\ \ln \frac{B}{B_0} &\approx -\beta r \end{aligned}$$

or

$$\frac{B}{B_0}$$

$$= \underbrace{\left(1 - e^{-\frac{r}{N}}\right)^2}_{\text{prob for viable doublet}} + \underbrace{\left(1 - e^{-\frac{r}{N}}\right)^2 e^{-\frac{\beta r}{N}}}_{\text{prob for dead doublet but surviving simplex}}$$

prob for viable doublet

prob for dead doublet but surviving simplex

for $\beta = 1$

$$\left(e^{-\frac{r}{N}} - 1\right) \left(1 - e^{-\frac{r}{N}}\right)^2 + 1$$

(O.K.)

for larger r and βr

$$\frac{B}{B_0} \approx 2e^{-\frac{r}{N}} + e^{-\frac{\beta r}{N}}$$

if $\beta < 1$

$$\ln \frac{B}{B_0} = -\frac{\beta r}{N}$$

if $\beta > 1$

$$\ln \frac{B}{B_0} = \ln 2 - \frac{r}{N}$$

$$N_1 + N_2 = N$$

$$\ln \frac{b_1}{b_0} \approx \underbrace{N_1 \ln 2 + N_2 \ln 3}_{\text{lost}} - \underbrace{(r_1 + r_2)}_{\substack{r \\ \text{lost on} \\ \text{chromosomes}}}$$

~~These chromosomes~~

These chromosomes

I and II equal III different

probable lost on I and II $\frac{r}{N}$
lost on III $\beta \frac{r}{N}$

Probab. that III survives is $e^{-\beta \frac{r}{N}}$

Probab that III ~~survives~~ is $(1 - e^{-\beta \frac{r}{N}})$

Probab that III is lost but (I+II) are viable
is $(1 - e^{-\beta \frac{r}{N}}) [1 - (1 - e^{-\frac{r}{N}})^2]$

Probab that (I+II+III) ~~are~~ is viable

$$\frac{B}{B_0} = e^{-\beta \frac{r}{N}} + (1 - e^{-\beta \frac{r}{N}}) [1 - (1 - e^{-\frac{r}{N}})^2]$$

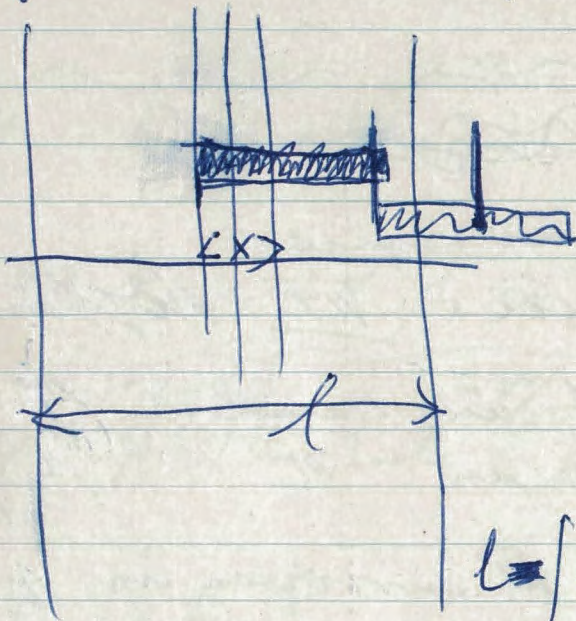
$$= \cancel{e^{-\beta \frac{r}{N}}} 1 - (1 - e^{-\frac{r}{N}})^2 + (1 - e^{-\frac{r}{N}})^2 e^{-\beta \frac{r}{N}}$$

side of strip because 0 of

$$\frac{d}{2} = 2a$$

$$d = 4a ;$$

lets strips make equal



$$l = \left[\frac{l}{2} - \frac{d}{2} + \frac{x}{2} \right] =$$

$$= x$$

$$\frac{l}{2} + \frac{d}{2} - \frac{x}{2} = x$$

$$\frac{3}{2}x = \frac{l+d}{2}$$

$$x = \frac{l+d}{3}$$

$$\frac{x}{2} = \frac{l+d}{6}$$

$$\frac{x}{2} = \frac{d}{2} - a \quad \text{means:}$$

$$\frac{l+d}{6} = \frac{d}{2} - a$$

$$= \frac{d}{2} - \left[d - \frac{l}{2} \right]$$

$$\frac{l+d}{6} = \frac{l}{2} - \frac{d}{2}$$

$$\frac{l+d}{3} = l - d$$

$$l+d = 3l - 3d$$

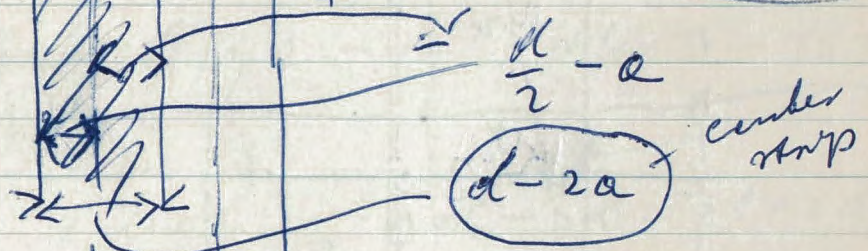
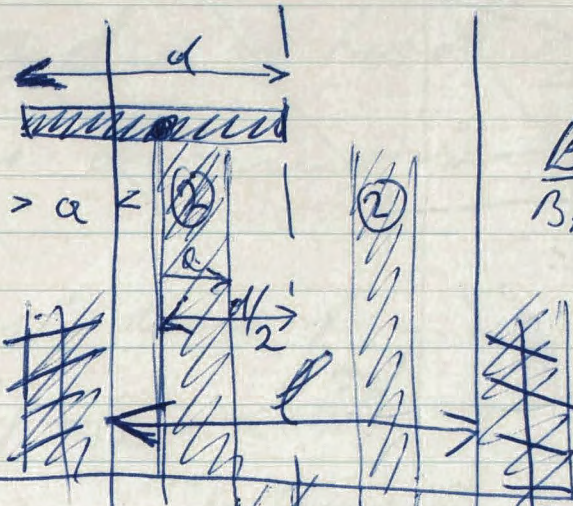
$$0 = 2l - 4d \quad d = \frac{l}{2}$$

Putland case (2 and 3 wts) / l

$$d - \frac{l}{2} = a$$

$$\frac{B}{B_1} = \left[1 - \left(1 - e^{-\frac{2a}{N}} \right)^2 \right]^N$$

$$\times \left[\quad \right]^N$$



$$d - \left[\frac{d}{2} - a \right]$$

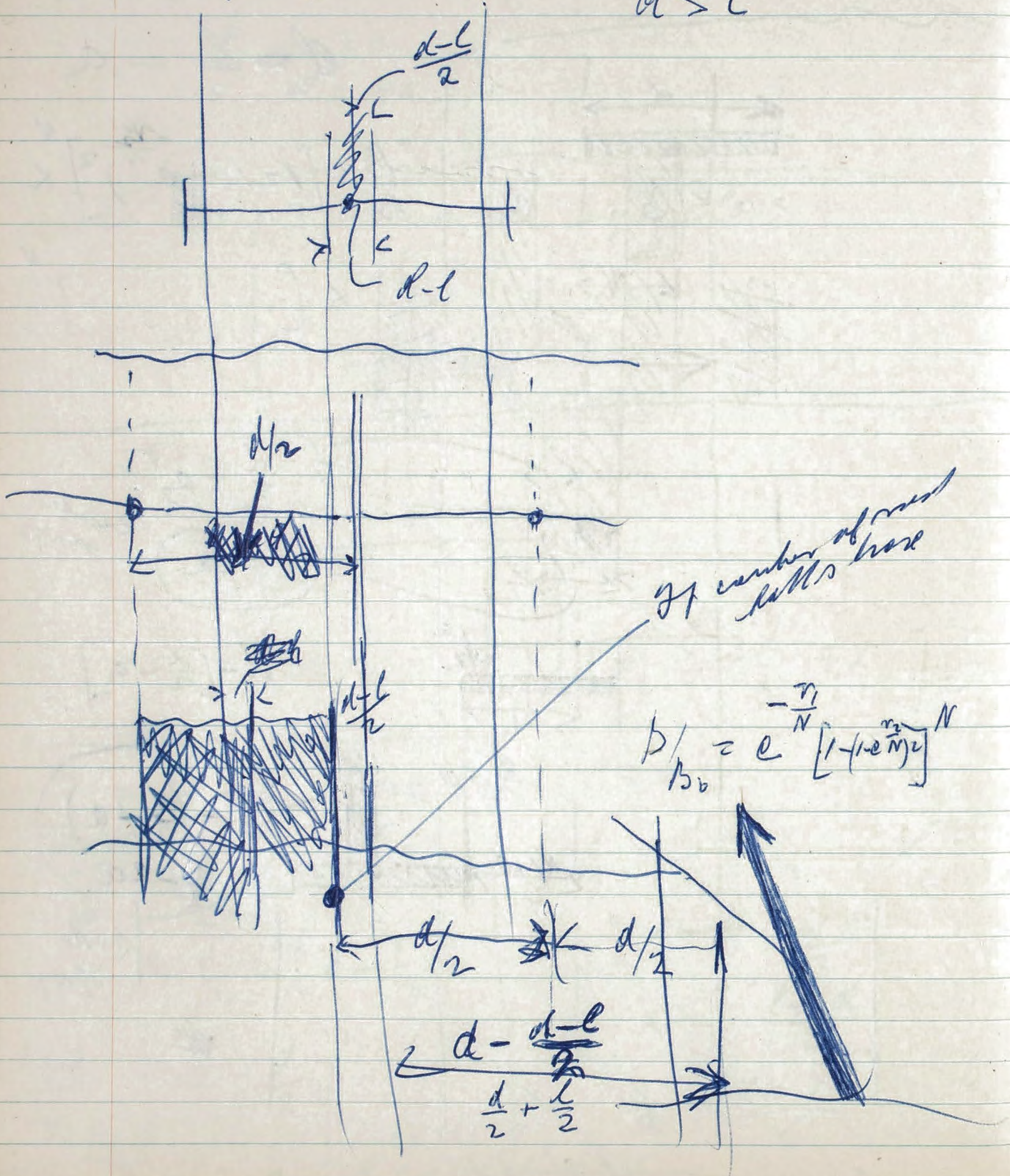
$$\frac{l}{2} - \left(\frac{d}{2} + a \right) = \frac{d}{2} - 2a$$

Hide strip

$$l + 2a$$

Case of one and two lvs

$$d > l$$



11

~~and~~
 ~~$2d - l + 3l + d =$~~
 ~~$3d$ is total of strips~~
 for three dimensional case
 total of strips

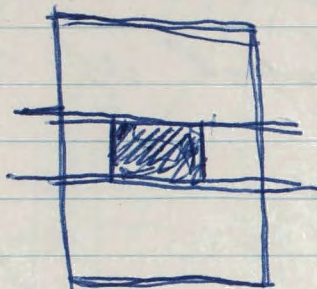
B will have form
 $\frac{B}{B_0} = \left[1 - \left(1 - e^{-\frac{r_2}{N}} \right)^2 \right]^N \left[1 - \left(1 - e^{-\frac{r_3}{N}} \right)^3 \right]^N \dots$
 for large r
 $\ln \frac{B}{B_0} \sim N \log 2 - r_2 + N \log 3 - r_3$

$d - \frac{l}{2} + \frac{l+d}{3} = \frac{6d - 3l + 2l + 2d}{6}$

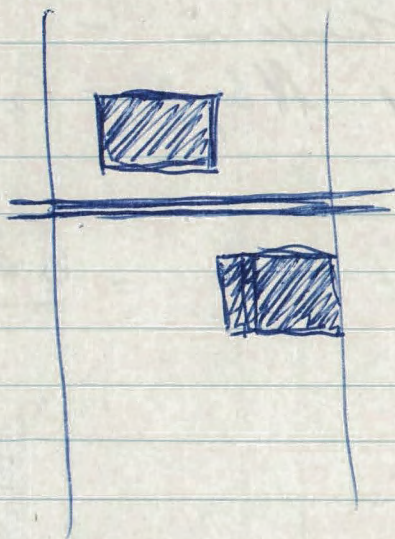
$\frac{8d - 2l}{6} > \frac{1}{3}l$
 $< 2l$

Machine

light flashes
at irregular
intervals



producing



counting

for ultimate slope in horizontal
case -

number of bits by center of
"rod" per cm be p
then ultimately everything may
be blocked except a small spot

~~number of bits~~
number of bits increasing by one
group than ~~the~~ d bits

New take rudimentary case of
1 and two bits -

center strip $d-l$

side strip $\frac{d}{2} - \frac{d-l}{2} + d-l$

$$d - \frac{d-l}{2} = \frac{d}{2} + \frac{d-l}{2} = d - \frac{l}{2}$$

~~one and two bit contour:~~

All wrong because certain
other combinations of two bits will
also work. -

unintentional improved

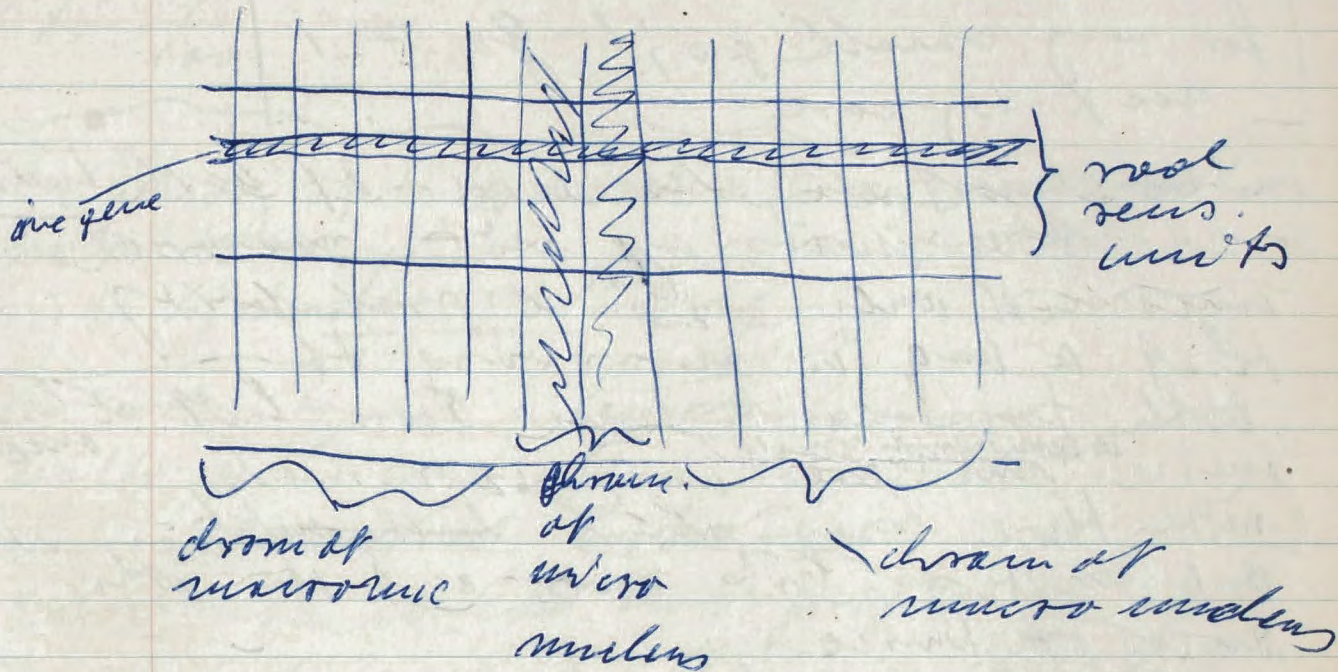
~~Experiments~~ assumption: within
a sphere of r lit with high
probability every thing destroyed
both the part of the gene but
that lies within the sphere and
neighbouring genes. However
rarely, say 1% once in 20 lits,
only the part of the gene but ~~is~~
~~the~~ which is lying within the sphere
is destroyed but not the neighbouring
genes. Let us assume here ~~at~~ the ^{next}
neighbouring genes are vital for survival.
If we now assume a "planned" case and
~~we now~~ have $\frac{r}{n} \approx 1$, one of the radiation
units ~~with~~ ~~the~~ ~~the~~, containing the
gene, will be destroyed with high
probability.

Zero point mutations $\frac{r}{N}$

Assumptions à la bellbruck

Assumptions: -

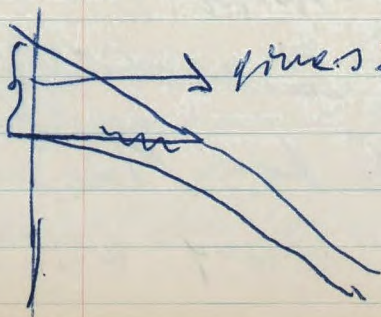
number of hits on 1 strand r



number of hits per radiological sensitive unit $\frac{r}{N}$

bellbruck Buren formula for survivors

$$\frac{B}{B_0} = \left(e^{-\frac{r}{N_1}} \right)^{N_1} \left(1 - \left(1 - e^{-\frac{r}{N_2}} \right) \right)^{N_2} \left(1 - \left(1 - e^{-\frac{r}{N_3}} \right) \right)^{N_3}$$



Let the probability that we find a mutation for a given dose be:

$$(1 - f_0)^2 \times 10^8 \text{ per survivor}$$

[for very small f_0 ; $1 - f_0 = 1$
see X-ray case]

since almost all are dead of both radiation units are hit we are concerned only since probability that a bug is surviving if both units have been hit at least once since the are only concerned with those cases where both units are hit once and the other once or twice

becomes exceedingly small if both radiation unit have been hit once and one of them is hit a second time we shall find that the number of mutants per survivor will increase with the dose proportionally to D^2 proportional to the square of the dose as long as

Facts from beemere:

50 recessive point mutants / 10^8
50000 dominant point mutants / 10^8
for 40000 R which kills B/r down
to $\frac{1}{100}$

The Theory of U.V. and X-ray induced mutations

1.) U.V. in diploid case

In order to get a ^{specific} mutation both twin radiation sensitive units must be hit. —

Each such unit has p+q genes.

Probability that a gene ~~is killed~~ ^{survives} is $\frac{1}{2}$ for all genes except the one in which we are interested. [if radiation unit is hit!]

In order to get a mutation all p genes must survive in both "chromosomes", but the one special gene must be killed in both

chromosomes; probability that the special gene ~~is killed~~ ^{survives} in one chromosome will be $\frac{1}{2}$

$$\frac{M}{B_0} = \frac{(1-p_0)^2 [1-(1-\alpha)]^2}{[1-(1-e^{-\frac{r}{N}})]^2} P (1-e^{-\frac{r}{N}})^2$$

or for ~~large~~ large $\frac{r}{N}$ denominator for

$$[2 e^{-\frac{r}{N}}]^2 = 4 e^{-\frac{2r}{N}}$$

This does not go with second power of dose!

try it differently: what is the

probability ~~to find~~ that a survivor has ~~one~~ ^{one} "surviving" special but on one chromosome and one surviving special but on ~~the~~ ^{its} twin chromosome?

The survivors fall into the following classes 1.) no hit on left special chrom. 2.) no hit on right special chromosome 3.) no hit on either, ~~the special both on~~

4. special hit on the left and special hit on the right

$$\frac{4}{1+2} = \frac{\text{const} \times [\text{small const}] (1 - e^{-\frac{r}{N}})^2}{2 e^{-\frac{r}{N}}} \left[\text{prob that hit only once} \right]$$

21

The probability of a hit
 on ~~the~~ radiation unit that
 (or)
 an appreciable fraction of
 the genes-survives is small.
 Let us see if this is true?

In order to get survival ~~of the lung~~
 must have all p genes survive
 in at least one of the two chromo-
 some,

i.e. a la Poisson
 probability that both genes die

$$(1-\alpha)^2$$

probability that lung survives

$$\left[1 - (1-\alpha)^2\right]^p$$

or if both radiation units are hit once

then probability of ^{alive} mutation

$$(1-g_0)^2 \left[1 - (1-\alpha)^2\right]^p = M$$

total number of mutations per survivor

~~$$\frac{M}{P_0} \approx \frac{(1-g_0)^2 \left[1 - (1-\alpha)^2\right]^p (1-e^{-\frac{r}{N}})^2 (1-(1-e^{-\frac{r}{N}}))^{N-2}}{\left[1 - (1-e^{-\frac{r}{N}})\right]^{N+1}}$$~~

Now let for a given dose for which
be: mutation/multivar = $f\left(\frac{r}{N}\right) = 1$

$$= (1 - f_0) 10^{-y}$$

or we derive (for large $\frac{r}{N}$)

$$(1 - f_0)^2 [1 - (1 - \alpha)^2]^p = (1 - f_0) 10^{-y}$$

$$\text{or } [1 - (1 - \alpha)^2]^p = \frac{1}{10^y}$$

$$\text{or } 1 - (1 - \alpha)^2 = \frac{1}{10^{y/p}} = A$$

$$\text{or } \alpha = 1 - \sqrt{1 - A}$$

$$\text{or } \text{since for } \frac{y}{p} \ll 1; A \approx 1 - \frac{2.3 y}{p}$$

$$\text{and } \alpha = 1 - \sqrt{\frac{2.3 y}{p}}$$

$$\text{for } p = 100; y = 6 \quad \alpha \approx 1 - \frac{1}{3} = \frac{2}{3}$$

prob that each hit only
once.

4

prob of hit $\times (1 - e^{-\frac{r}{N}})$

$$(1 - f_0) [1 - (1 - \alpha)^2] \left[\frac{r}{N} e^{-\frac{r}{N}} \right]^2 [1 - (1 - e^{-\frac{r}{N}})]^{N-1}$$

$$[1 - \dots]^{N-1} + [1 - (1 - e^{-\frac{r}{N}})]^2]^N$$

for large $\frac{r}{N}$ ~~for small $\frac{r}{N}$~~

$$= (1 - f_0)^2 [1 - (1 - \alpha)^2]^2 \left(\frac{r}{N} \right)^2 \left(e^{-\frac{r}{N}} \right)^2$$

$$[1 - (1 - e^{-\frac{r}{N}})]^2$$

$e^{-\frac{r}{N}}$

for large $\frac{r}{N}$

$$\approx \frac{(1 - f_0)^2 [1 - (1 - \alpha)^2]^2 \left(\frac{r}{N} \right)^2}{2 \left[e^{-\frac{r}{N}} \right]^2}$$

more than
one

We ~~if you~~ neglect some multiplications
of one at the two subdomains with ~~at~~ both

$$(1 - q_0)^k \left[1 - (1 - \alpha)^k \right]^p \left(\frac{m}{N} \right)^k$$

$$p = 100$$

$$\alpha = \frac{2}{3}$$

2 standard uses

$$(1 - \frac{2}{3})^k \quad \cancel{\left(\frac{2}{3} \right)^k} \quad \left(1 - \frac{1}{3} \right)^k \quad p = 2^k$$

$$k = 1$$

$$\left(\frac{2}{3} \right)^k$$

$$k = 2$$

$$\left(\frac{8}{9} \right)^{100}$$

$$e^{\frac{100}{9}}$$

$$\left[1 - \frac{1}{9} \right]^{\frac{100}{9}}$$

$$\left[1 - \frac{1}{27} \right]$$

$$e^{-11}$$

$$\boxed{1000}$$

$$\textcircled{310^{-5}}$$

$$\frac{3000}{1000}$$

$$e^{-10}$$

$$10 \frac{2.80}{2.13}$$

$$e$$

$$e^{-1}$$

fresh start on V.V. numbers
 assume $\frac{r_0}{N}$ large $\frac{r_0}{N}$ number of

numbers M_0 , we now increase

r by Δr what is $\frac{dM}{dr} = \frac{M'S - MS'}{S^2}$

$$S = S_0 e^{-r}$$

$$S' = -S_0 e^{-r}$$

$$\Delta M = \frac{1}{2} (\text{factor})^2 \frac{r}{N} e^{-r/N} \frac{\Delta r}{N} S'$$

$$\frac{\Delta M}{\Delta r} = \frac{1}{N} S \times 2 \times (\text{factor})^2 \frac{r}{N} e^{-r/N} - M e^{-r}$$

$$\frac{dM}{dr} = \frac{1}{N} S^2 \times 2 \times (\text{factor})^2 \frac{r}{N} e^{-r/N} + \frac{M}{S} e^{-r}$$

$$\frac{dM}{dx} = \dots$$

$$\frac{r}{N} = x$$

$$\frac{dr}{N} = dx$$

$$dr = N dx$$

$$2 \times \frac{1}{N} S^2 \times (\text{factor})^2 \frac{r}{N} e^{-r/N} = x^2 e^{-x}$$

New for X-rays:

H

probability of mutant

$$\alpha^p (1-q_0) \frac{r}{N} e^{-r/N} \left(e^{-\frac{r}{N}}\right)^{N-1}$$

~~XXXXXXXXXX~~

$$\left(e^{-\frac{r}{N}}\right)^N$$

V.V. case considered

no ~~of~~ ~~(1-q_0)~~ ~~is~~ let q_0 be very small
 $(1-q_0) = 1$

then we have following higher terms

$$\left(\frac{r}{N}\right)^2 \frac{p}{2} (1 - (1-\alpha)^2) \left[\frac{r}{N}\right]^2 +$$

$$\frac{p}{2} \left[1 - (1 - e^{-\frac{r}{N}})^2\right]$$

$$+ \left[1 - (1-\alpha)(1-\alpha^2)\right] \frac{p}{N} \left(\frac{r}{N}\right)^2 \frac{p}{2}$$

$$+ \left[1 - (1-\alpha^2)(1-\alpha^2)\right] \left(\frac{r}{N}\right)^2 \left(\frac{r}{N}\right)^2 \frac{1}{2!} \frac{1}{2!} +$$

+ ...

A Cleanup in Traits

Northwood Diploid Theory

assumption + N mutation sensitive unit, each "100" genes probability μ of one gene dying in case of hit. —

$$\frac{P}{P_0} = \text{[scribbled out equation]}$$

probability of survival of one gene if μ hits $\frac{-\mu r}{N}$

of death of one gene $1 - e^{-\frac{\mu r}{N}}$

of death of both genes

$$[1 - (1 - e^{-\frac{\mu r}{N}})]^2$$

of ~~rest~~ remaining viable

$$1 - (1 - e^{-\frac{\mu r}{N}})^2$$

of all remaining viable

$$\left(1 - [1 - e^{-\frac{\mu r}{N}}]^2\right)^N$$

for very high r

assumption of independence of genes!!!

d - days

H

$$\frac{M}{S} = d^p (1 - q_0) \frac{r}{N} \quad \text{for } \frac{r}{N} \text{ close to } 1$$

40000 R units give $\frac{1}{100}$ survivor

or $r = \frac{40000}{100} = 400$ close
for $r = 1$ we have 4000 R units

$$5 \cdot 10^{-5} = d^p (1 - q_0) \frac{400}{N}$$

$$10^{-5} \approx d^p (1 - q_0) \frac{1}{N}$$

10000

20000

(-5)
210

for $N = 10$

$$q_0 = 0$$

$$10^{-4} = d^p$$

$$p = 100$$

$$d^{100} = 10^{-4}$$

$$d = 10^{-\frac{4}{100}}$$

$$10^{\frac{1}{100}} e^{-\frac{2.3}{100}} \approx 1 + \frac{2.3}{100}$$

$$= e^{-\frac{2.3}{25}} \approx e^{-\frac{1}{10}}$$

$$= 1 - \frac{1}{10}$$

$$d = 0.9$$

or corresponding probability of death
only 0.1

putting now $1 - q_0 = \frac{1}{10} - \frac{2.3}{33}$

$$10^{-3} = d^p \quad \left| \quad e^{-\frac{2.3}{33}} = 1 - \frac{2.3}{33}$$

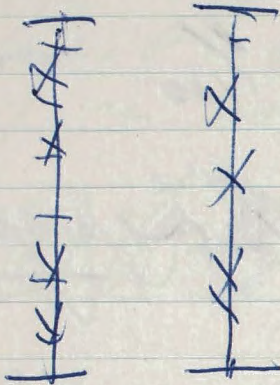
(1/14)

2.3/33

Products at 1 last left 1 last right

$$P(i) \times P(i) +$$

$P(i)$



$P(k)$

Matrix Problem best way!

4

$$\left[2 e^{-\frac{\alpha r}{N}} \right]^{pN}$$

H

$$2^{pN} e^{-p\alpha r}$$

$$\ln \frac{B}{B_0} = pN \ln 2 - p\alpha r$$

But this does not help to get a larger N except that we made assumption of independence of cuts in same or different genes in same restriction unit.

Try better:

"viability in one section":
 probability of both the main sections



M

Alarvic 30 to 40 cents
per lbs Polyethylene
extruded
make 40 to 50 cents

Span sheet per ton
100 dollars

Aluminum sheet 25c/lb
 $\frac{10}{1000}$ of inch

25 cents per 1000 entire
sheet

and in bulk per ton

40 cents per 1000 entire sheet

Buy Algae for humans \$150 / ton
for animal \$60 / ton

10,000 foot candles Arizona H
desert at noon

Empire & Quarterly last issue
The American Scientist
Rumrystan Kandel.

Wisconsin

panels

Burris [Agric. Machinery]

(intrusion) Wisconsin at
Blue Green

and California type work

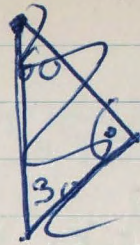
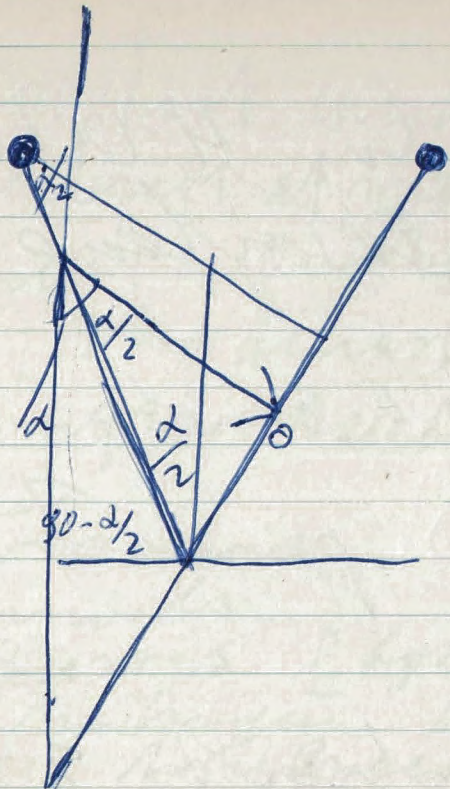
Manufacturer: Wolfe Scoop
production, production
of Blue Green

Illinois \$200 per acre

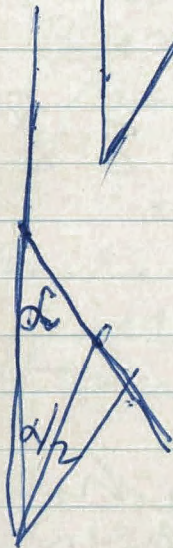
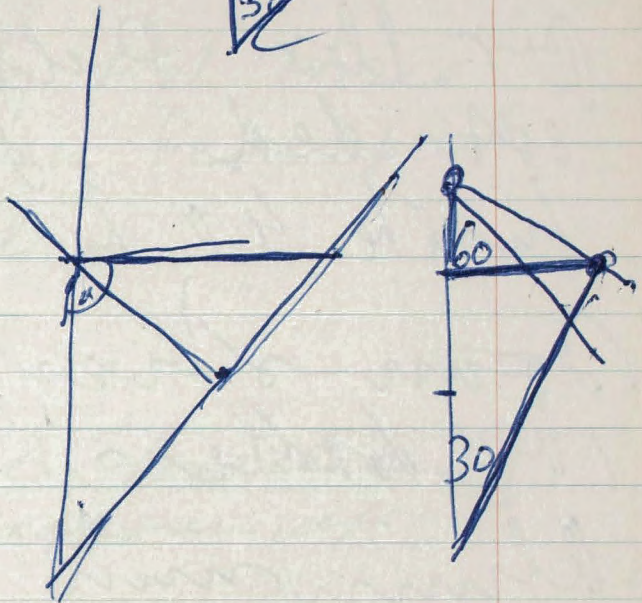
Antique car in California \$500

Chlorine with 10 - 50 lbs dry
\$5000 per acre
merchandise

including 30% of cost
percentage 10%



14



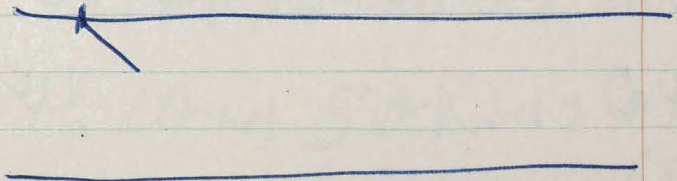
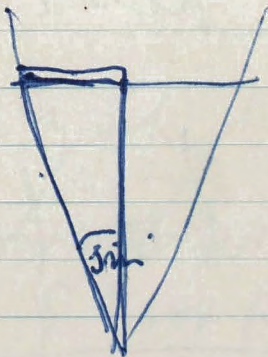
$$L + \alpha/2 \neq 90^\circ$$

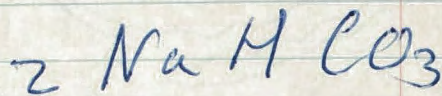
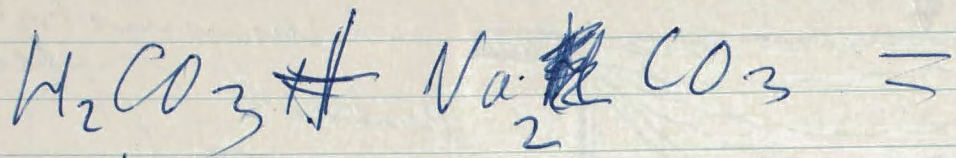
$$\frac{3}{2} \alpha = \text{req } 90$$

60°

$$\sin \frac{\alpha}{2}$$

$$\sin 30$$



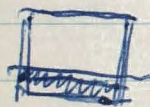


$$C_1 \times C_2 = C_3$$

Line graph

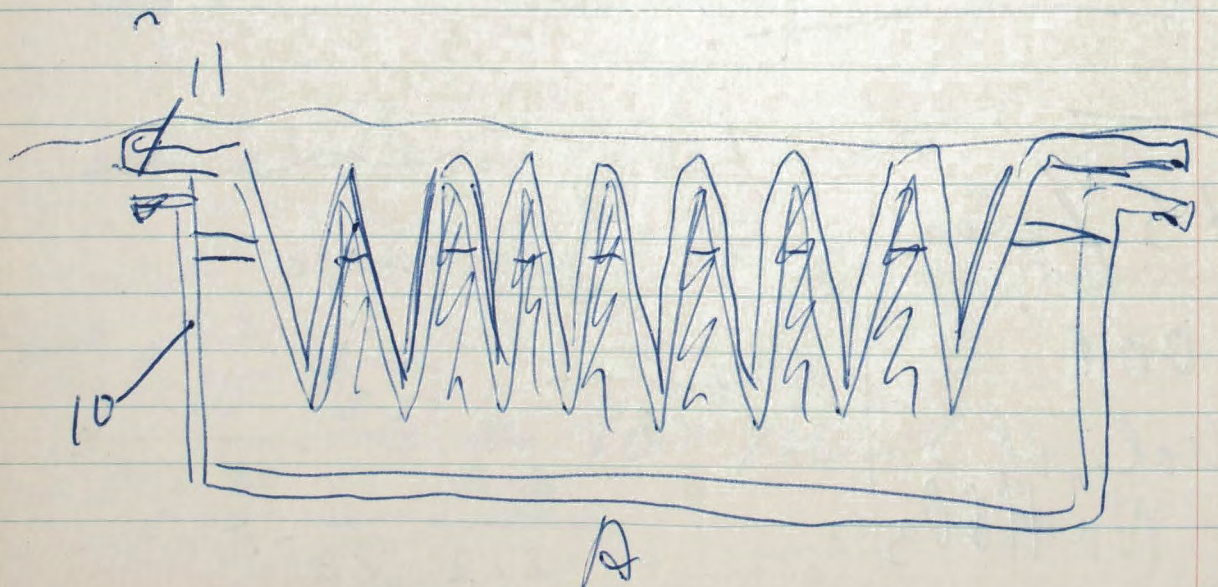
$\frac{C_3}{C_2}$

3.5 ⁻³ 10³ ca/ae



$\frac{3.5}{1000}$ cm

$\frac{35}{1000}$ of m/m



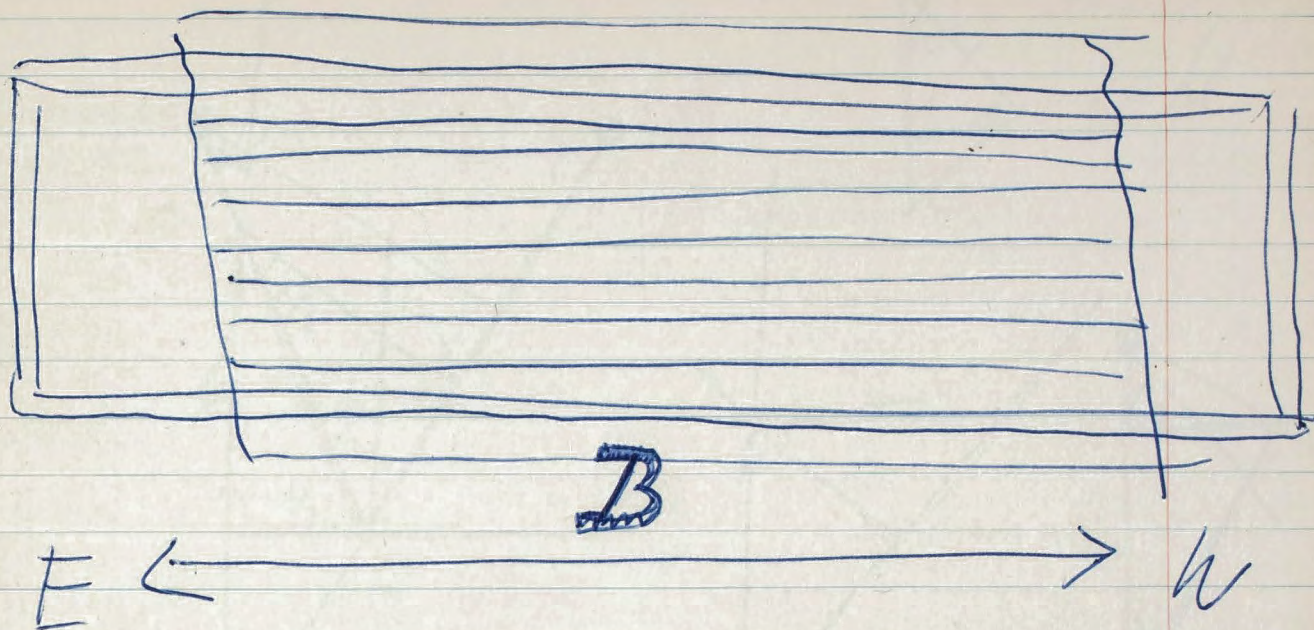
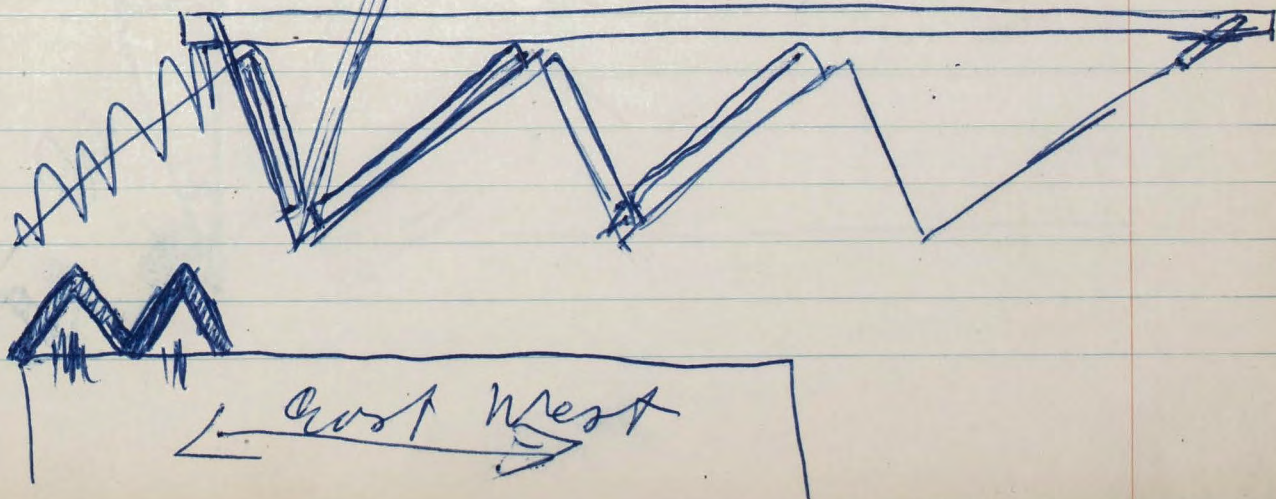
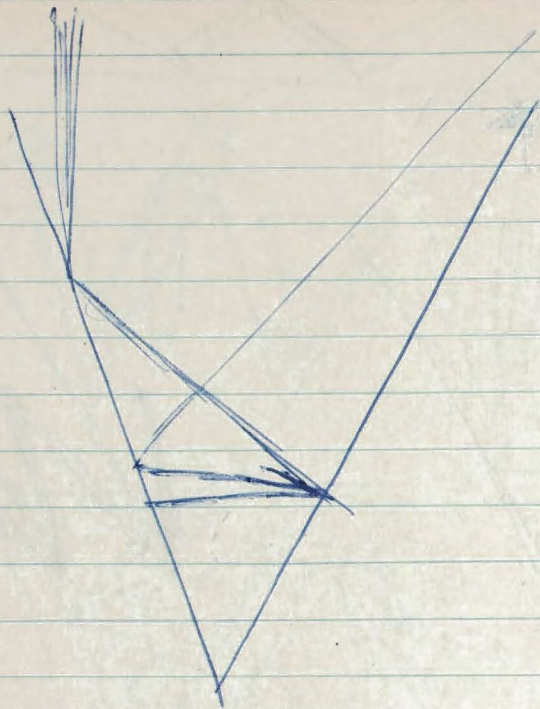


Fig 3

Sun at noon on March -
- summer





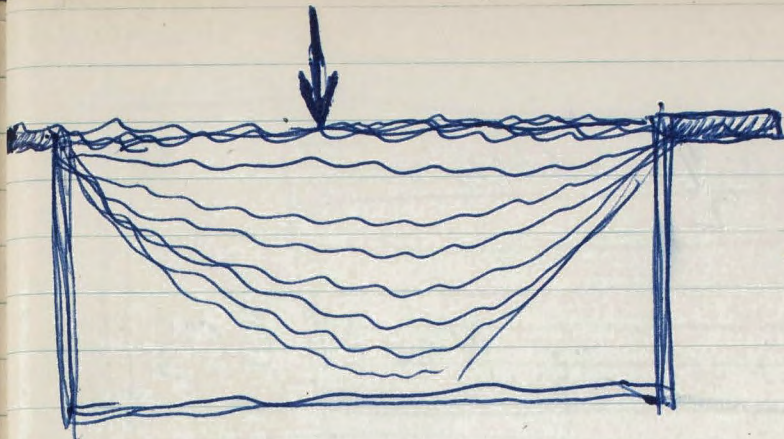
at 5% CO_2 $\frac{1}{1000}$ of NaHCO_3
 at 3×10^{-4} pH 6

$\frac{3}{100}$ ~~per~~ % CO_2 $\frac{3}{10^5}$ Mmol of NaHCO_3

1000 cc CO_2 gas / liter water

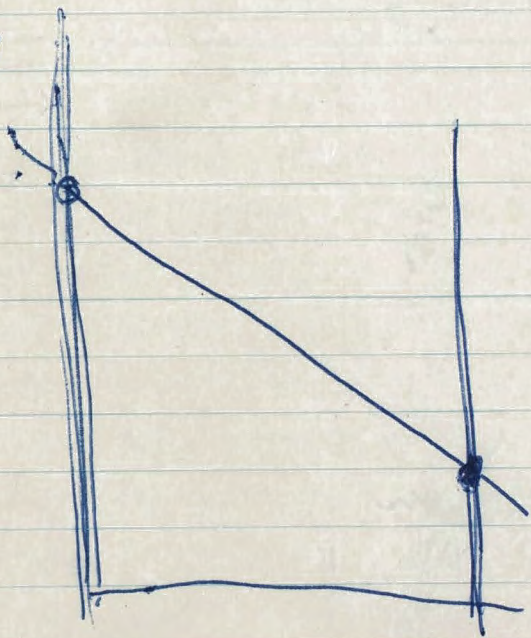
is $\frac{1}{22}$ of Mmol at this pressure

$$\frac{3 \times 10^{-4}}{22} = \frac{3}{2.2} 10^{-5} \text{ Mmol}$$



page 25
 manometric techniques
 Ambient 6

2567



1. ~~NaHCO₃~~ CO₂

5 to 7

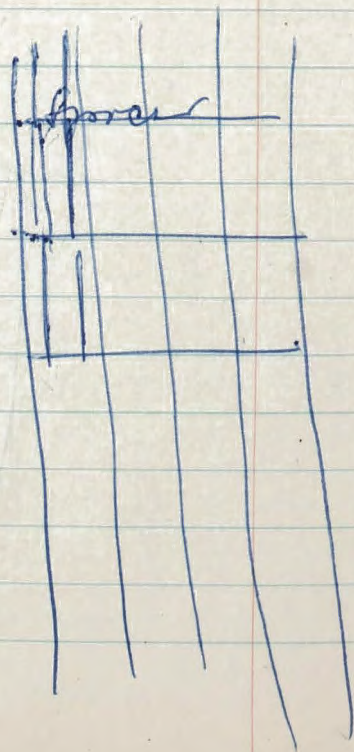
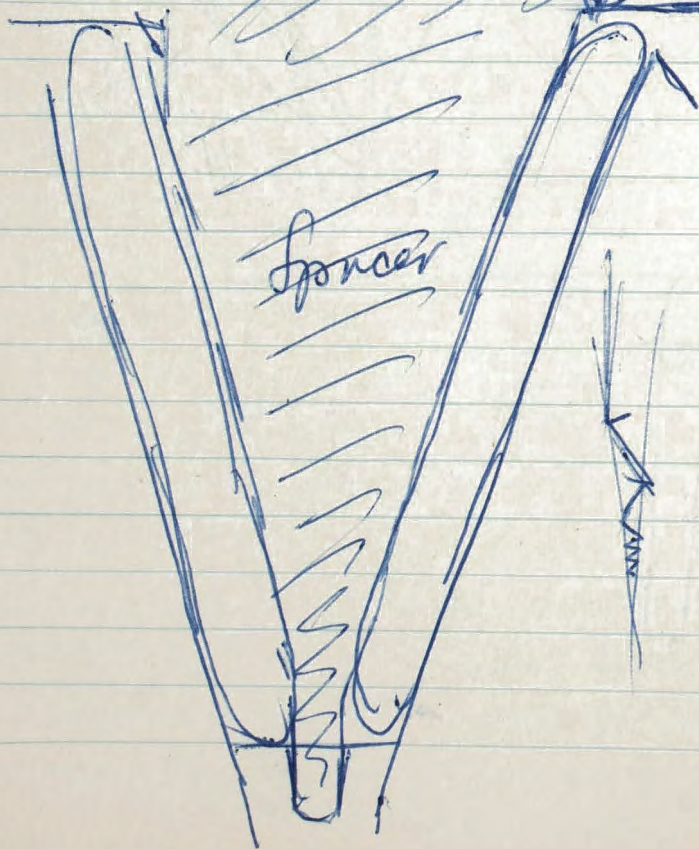
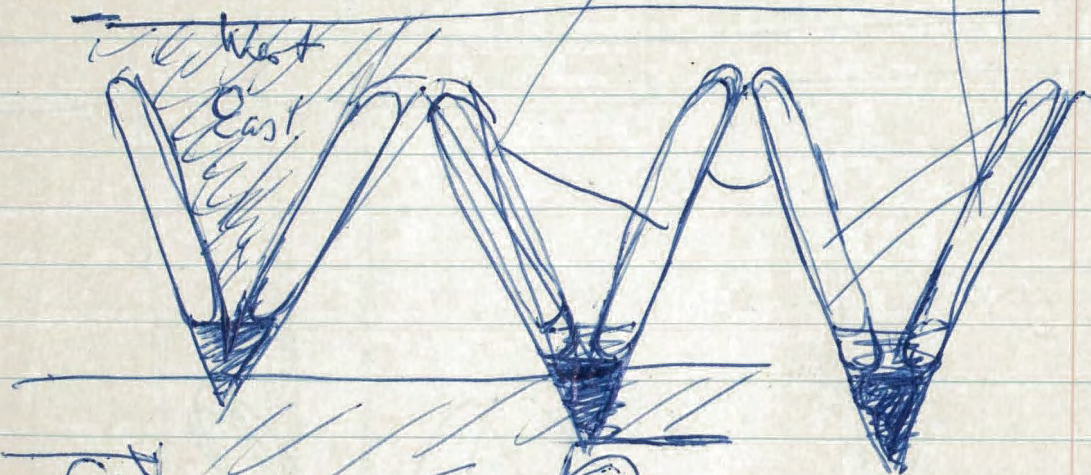
4.5 in test 2

~~10~~ $\frac{3}{1000}$ % CO₂ in air

$\frac{1}{200}$ mol

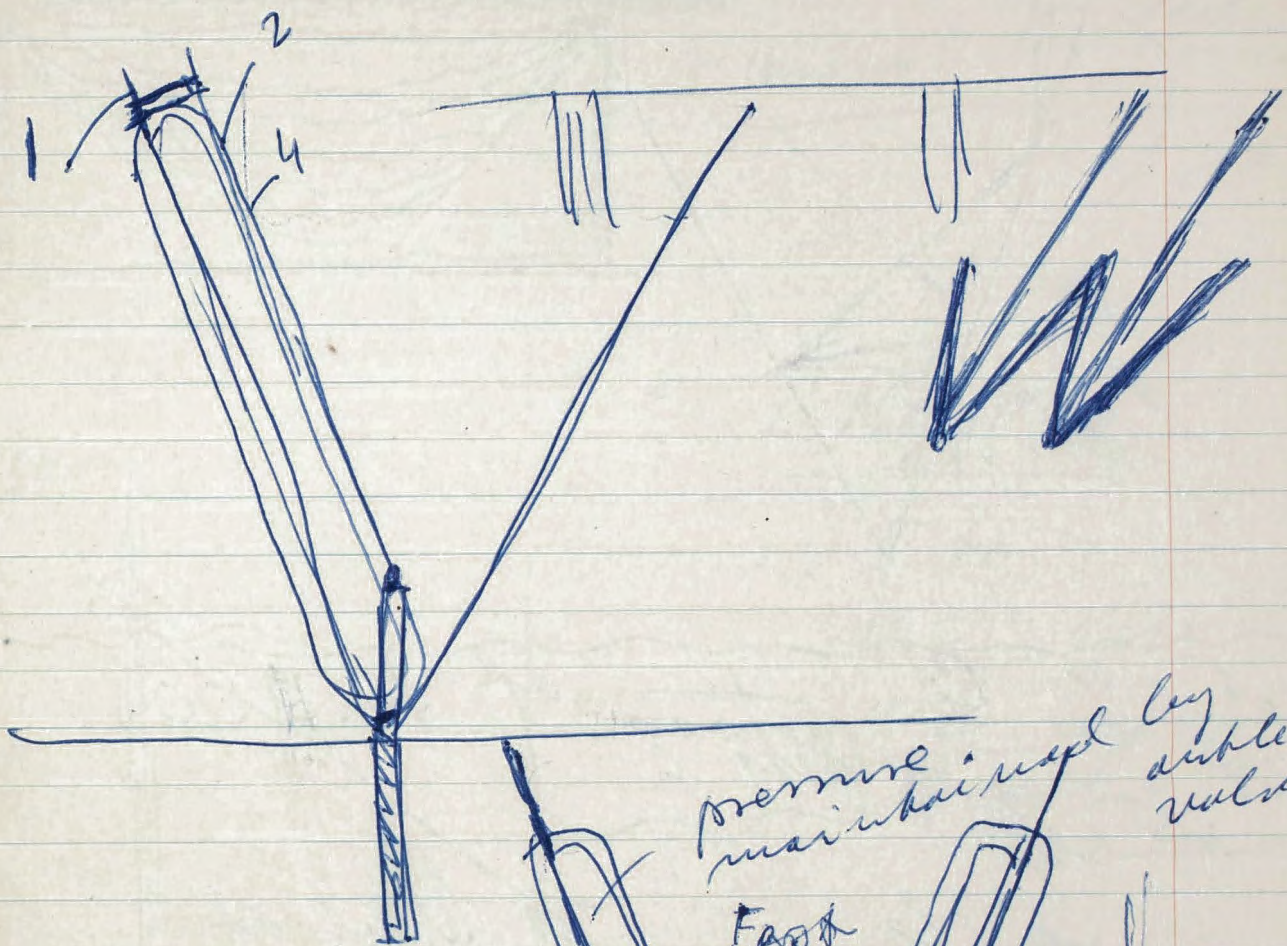
at 5% CO₂
 0.001
 mol of
 NaHCO₃
 PH 6.1

~~10000~~ 10000
 $\frac{3}{104}$ $\frac{1}{22}$ of a mol
 $\frac{3}{2.2} \cdot 10^{-5}$



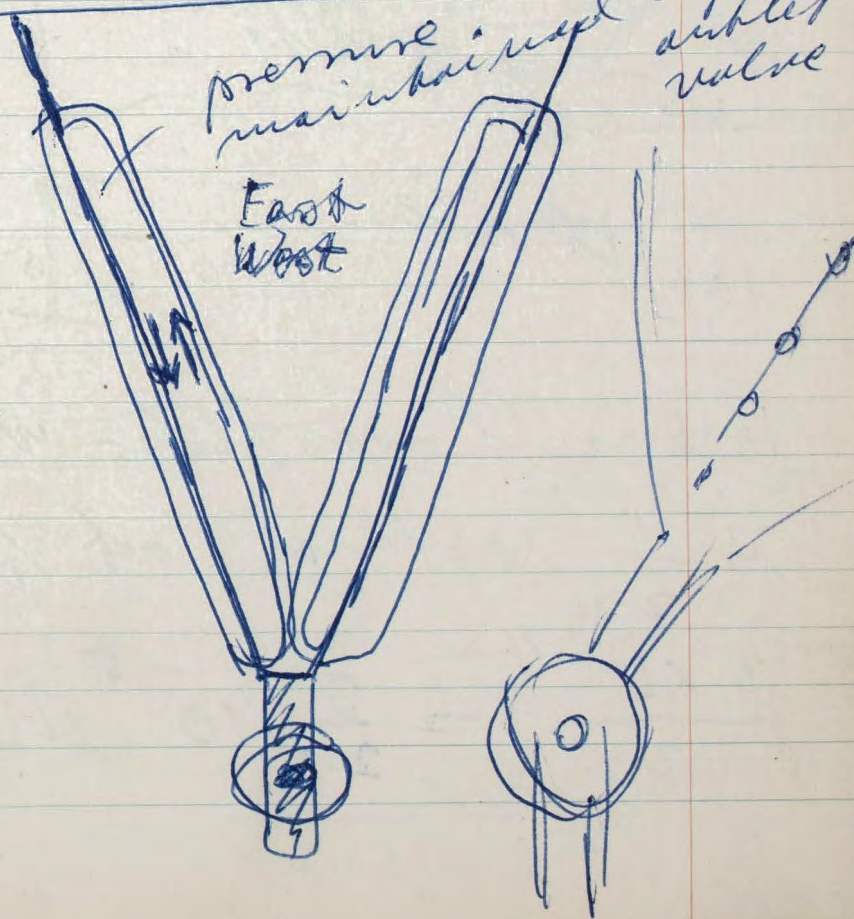
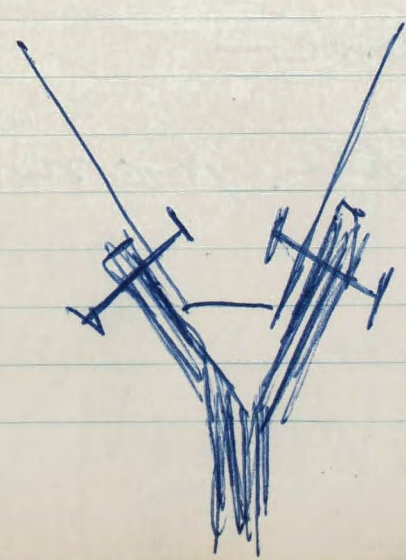
E

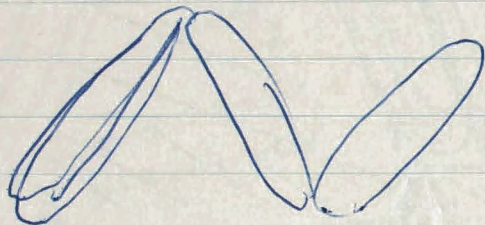
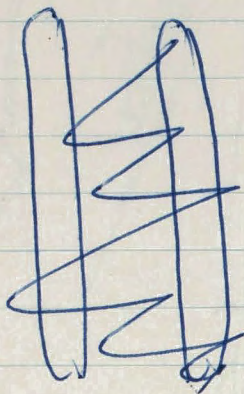
W



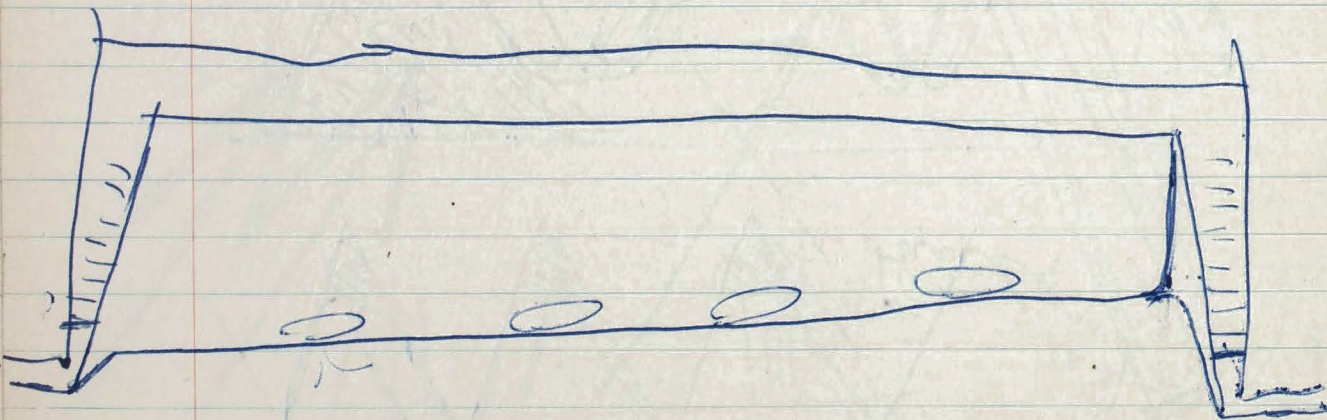
pressure
main
by
airlet
valve

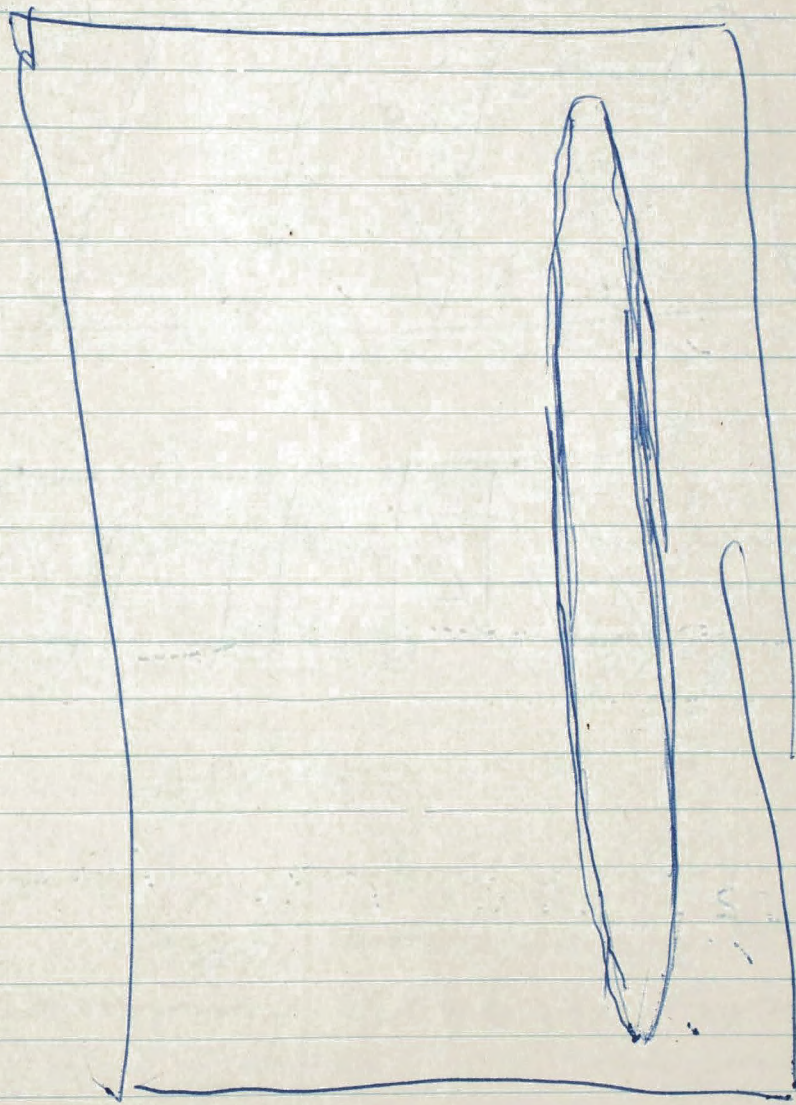
East
West





5





Sunlight

Chlorella

Wick Myers Journal of Gen
Biol Texas A&M

1 sunlight requires by animals

10 use $\frac{1}{50}$ of sunlight

Reiche & Gaffron chopped light

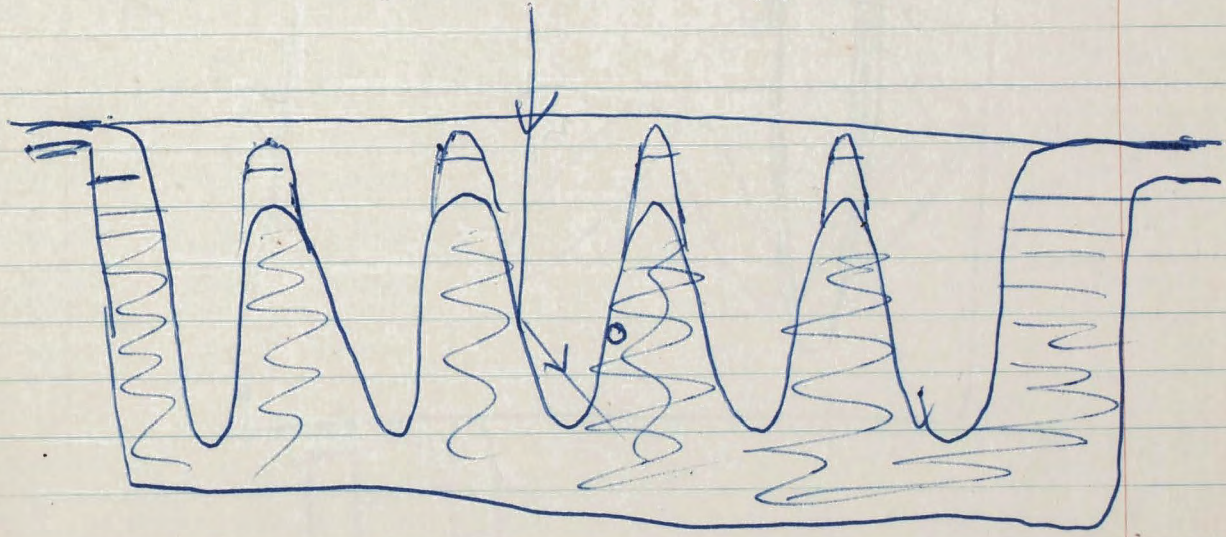
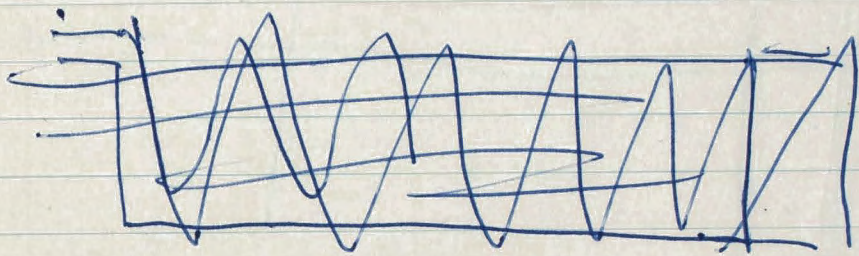
CO₂ fixation over Na₂CO₃-NaOH
Unbrat. ~~staphylococcus~~ Mononucleic
Acids

Maximum 1 hr 2 cc packed per liter
cell vol 3×10^{-11} cc

1 cm $\frac{1}{2}$ volume for light

Trace elements

Na₂CO₃ (at pH 4) or glycine buffer



Read Cowdry Problems of aging
1844².

Sakshansky functions and the
origin of species

For transcription:

Ann. of Exp. Medicine Apr/49

H. Taylor

Musford and Austroan

Kelner A.

90. Duff Str.

Wabertown Mass.

Laudauer

12/1/20

E 12

Algae

1950

